

*Review article*

Effect of Temperature, pH, Salinity and Dissolved Oxygen on Fishes

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manan.chatha@iub.edu.pk**Abstract**

Environmental factors, including temperature, pH, salinity, and dissolved oxygen, are paramount in shaping fish physiology, behavior, and survival. Fish, being highly responsive to these environmental shifts, undergo profound changes in metabolism, growth, and overall performance. Specifically, temperature variations can have acute or long-term effects, pH changes disrupt ion balance and respiratory efficiency, salinity affects osmoregulation and ion dynamics, and dissolved oxygen levels are fundamental for respiration and metabolic health. Understanding these intricacies is not just academic; it's crucial for fisheries management, conservation strategies, and anticipating the ramifications of broader environmental alterations. This review offers an in-depth analysis of these environmental impacts on fish, underscoring the significance of each factor in their physiology, adaptive behaviors, and ecological context.

Keywords: pH, salinity, dissolved oxygen, fish

1. Introduction

Fish are cold-blooded aquatic animals that depend highly on their surrounding environment for survival and growth. Environmental factors such as temperature, pH, salinity, and dissolved oxygen are critical in fish's life cycle and physiological processes. These environmental factors can significantly impact fish physiology, behavior, and population dynamics. Therefore, understanding the effects of these factors on fish is crucial for successful fisheries management and conservation [1]. Understanding these effects is not just about the well-being of individual species; it's about the bigger picture. Changes in fish behavior, reproduction, growth, and survival can alter predator-prey dynamics, nutrient cycling, and overall biodiversity within aquatic habitats [2]. Temperature influences various aspects of fish biology, including growth rates, metabolic rates, reproduction, and overall performance. Both acute and chronic temperature changes can have significant effects on fish physiology and behavior. Higher temperatures can accelerate metabolic

processes, increase energy requirements, and lead to oxygen limitation, while lower temperatures can decrease metabolic rates, impair enzyme activities, and affect immune function. Temperature fluctuations can also influence the distribution and abundance of fish species, with potential implications for ecosystem dynamics [3].

pH represents the acidity or alkalinity of the water, affecting the ion balance and acid-base regulation of fish. Changes in pH can alter the availability and toxicity of certain substances, impacting fish at cellular and systemic levels. Acidic conditions (low pH) can disrupt ion regulation, impair enzyme activities, and compromise reproduction. Conversely, alkaline conditions (high pH) can lead to the accumulation of toxic substances, affect respiratory efficiency, and induce stress responses in fish [4].

Salinity, the concentration of dissolved salts in water, is a critical factor influencing fish physiology and osmoregulation. Fish species exhibit varying tolerances to salinity levels, and changes in salinity can disrupt ion balance, osmotic regulation, and reproductive processes. While euryhaline species can

tolerate a wide range of salinities, stenohaline species have specific salinity requirements, making them more susceptible to habitat alterations and salinity fluctuations [5]. Dissolved oxygen (DO) is essential for fish respiration and aerobic metabolism. Insufficient DO levels, often associated with eutrophication or pollutant inputs, can lead to hypoxic or anoxic conditions in water bodies, posing serious threats to fish survival and fitness. Low oxygen levels can impair growth, reproduction, and immune function, and may result in population declines or even mass mortality events [6].

Temperature, pH, salinity, and dissolved oxygen are critical environmental factors affecting fish physiology, behavior, and population dynamics. Changes in these environmental factors can significantly impact fish growth, reproduction, and survival. Therefore, understanding the effects of these factors on fish is crucial for successful fisheries management and conservation [7].

2. Temperature:

Fish exhibit a strong relationship between temperature and metabolic rate. As temperature increases, the metabolic rate generally rises, increasing energy requirements. Fish allocate energy differently under different temperature regimes, prioritizing growth and reproduction in optimal temperatures. However, under extreme temperatures, metabolic adjustments may occur to conserve energy and maintain essential physiological functions [8].

Temperature directly affects oxygen consumption rates in fish. Fish generally have higher oxygen demands as temperature increases due to increased metabolic activity (Table 1). Thermal tolerance limits define the upper and lower temperature thresholds at which fish can function physiologically. These limits are essential in understanding the potential impacts of temperature changes on fish survival and fitness [9].

Temperature influences the growth and development of fish. Higher temperatures generally accelerate growth rates, while lower temperatures can lead to reduced growth. Temperature also affects size at maturity, with fish in warmer environments typically maturing smaller than those in more relaxed

environments fish adjust their circulatory and cardiovascular systems in response to temperature changes. Higher temperatures can increase heart rate and blood flow, facilitating oxygen delivery to tissues. Conversely, lower temperatures can reduce heart rate and blood flow, affecting overall physiological function [10].

Temperature variations influence the immune system response of fish. Temperature changes can modulate immune system activity, affecting the efficiency of immune responses to pathogens and diseases. Elevated temperatures may enhance immune response, while lower temperatures can impair immune function, making fish more susceptible to infections [11].

Temperature plays a crucial role in determining the timing of fish spawning. Warmer temperatures often accelerate spawning, leading to earlier reproductive events. Fecundity, the number of eggs fish produce, can also be influenced by temperature, with higher temperatures generally associated with increased fecundity [12]. Temperature influences gonadal development in fish. Warmer temperatures can accelerate gonadal maturation, leading to earlier sexual development and breeding. Additionally, the temperature can impact the sex ratio of fish populations, with some species exhibiting temperature-dependent sex determination [13].

Temperature variations affect the survival and recruitment of fish larvae. Larvae have specific thermal requirements for survival and growth; temperature changes can impact larval development and survival rates. Temperature fluctuations may lead to shifts in recruitment patterns, potentially influencing population dynamics [14]. Temperature changes can influence the behavior of fish during courtship and mating. Fish may alter their reproductive behaviors, such as courtship displays and mate choice, in response to temperature variations. These behavioral adjustments can affect reproductive success and the genetic composition of fish populations. Each fish species has a unique thermal optima range, representing the temperature range in which they thrive. Temperature tolerance limits define the upper and lower thresholds beyond which the species can not survive.

Table 1. Effects of different temperature on fish.

Temperature	Effect on Fish	Reference
> 20°C	Optimal for most freshwater fish species, allows for maximum growth and reproduction	[18,19]
> 25°C	Increases metabolic rate, but can cause stress and decrease oxygen-carrying capacity of blood	[20]
> 28°C	Can cause cellular damage and lead to organ failure, decreased growth and reproduction	[21]
> 30°C	Can cause severe stress and mortality, especially in coldwater species	[22]
> 35°C	Lethal to most fish species within hours	[23]
< 5°C	Decreases metabolic rate and activity, can cause stress and decreased growth and reproduction	[24]
< 0°C	Can cause freezing of bodily fluids and tissue damage, decreased growth and reproduction	[25]
-1 to 4°C	Optimal for some coldwater fish species, allows for maximum growth and reproduction	[26]
5-10°C	Optimal for some coolwater fish species, allows for maximum growth and reproduction	[27]

Temperature changes can result in shifts in species distributions as they seek environments within their thermal tolerance range. We explore how certain fish species uniquely respond to temperature fluctuations, shedding light on the intricate dynamics of temperature-fish interactions.

1. Salmon (*Salmo salar*): As ectothermic organisms, salmon are particularly susceptible to temperature changes [15]. optimal growth occurs between 10°C and 14°C, while temperatures exceeding 20°C can be lethal due to increased metabolic stress and lowered dissolved oxygen levels. Spawning patterns of salmon are also temperature-dependent, with shifts potentially affecting their reproductive success.

2. Zebrafish (*Danio rerio*): Widely used as a model organism, zebrafish exhibit significant developmental changes when exposed to varying temperatures [16]. Typically, development rates increase with temperatures ranging from 24°C to 28°C but decrease or halt altogether outside this range

3. Clownfish (Amphiprioninae): Living in symbiosis with sea

anemones, clownfish are found in warmer waters. However, prolonged exposure to temperatures above 30°C affects their symbiotic relationships and may lead to bleaching events [17].

Temperature plays a crucial role in fish specie's physiological processes, growth, reproduction, and overall well-being. Different temperature ranges affect fish differently, influencing their metabolism, oxygen-carrying capacity, organ function, growth, and reproduction. Temperature above 20°C is optimal for most freshwater fish species, as it allows maximum growth and reproduction. As the temperature increases above 25°C, fish experience an elevation in metabolic rate. However, high temperatures can also cause stress and decrease the oxygen-carrying capacity of the blood, impacting fish physiology [28]. When exposed to temperatures above 28°C, fish may suffer cellular damage and organ failure, leading to decreased growth and reproduction [29].

Table 2. Effect of different pH on Fish

pH Level	Effect on Fish	Reference
6.5-9.0	Optimal for most freshwater fish species, allows for maximum growth and reproduction	[45]
< 6.5	Stressful for most fish species, can cause reduced growth and reproduction	[45]
< 6.0	Life-threatening to most fish species, can cause death if exposure is prolonged	[46]
> 9.0	Stressful for most fish species, can cause reduced growth and reproduction	[47]
> 9.5	Life-threatening to most fish species, can cause death if exposure is prolonged	[48]
< 4.0	Life-threatening to most fish species, can cause death if exposure is prolonged	[49]
6.5-8.5	Optimal for most saltwater fish species, allows for maximum growth and reproduction	[50]
< 6.5	Stressful for most saltwater fish species, can cause reduced growth and reproduction	[51]
6.5-9.0	Optimal for most freshwater fish species, allows for maximum growth and reproduction	[52]
< 6.5	Stressful for most fish species, can cause reduced growth and reproduction	[53]
< 6.0	Life-threatening to most fish species, can cause death if exposure is prolonged	[52]
> 9.0	Stressful for most fish species, can cause reduced growth and reproduction	[55]
> 9.5	Life-threatening to most fish species, can cause death if exposure is prolonged	[56]
< 4.0	Life-threatening to most fish species, can cause death if exposure is prolonged	[55]
6.5-8.5	Optimal for most saltwater fish species, allows for maximum growth and reproduction	[58]
< 6.5	Stressful for most saltwater fish species, can cause reduced growth and reproduction	[59]
4.0-6.0	Tolerable range for some fish species, may exhibit reduced reproductive capacity	[60]
8.5-9.5	Tolerable range for some fish species, may experience mild stress	[61]
> 10.0	Life-threatening to most fish species, can cause severe physiological disturbances	[62]

Temperatures exceeding 30°C can result in severe stress and mortality, particularly in Cold water fish species [30]. Extremely high temperatures, such as those exceeding 35°C, can be lethal to most fish species within hours [31]. At temperatures below five °C, fish experience decreased metabolic rate and activity. This can lead to stress and reduced growth and reproduction [32]. Subzero temperatures can cause freezing of bodily fluids and tissue damage, resulting in decreased growth and reproduction [33].

3. pH

Fish possess sophisticated mechanisms to maintain their internal acid-base balance, which is crucial for proper physiological functioning. pH changes in their environment can disturb this balance, leading to disruptions in acid-base regulation. Acidic or alkaline conditions can challenge fish homeostasis, affecting ion regulation, enzyme activity, and metabolic processes pH alterations affect the respiratory system of fish, impacting oxygen transport and delivery to tissues.

Enzymes, being biological catalysts, have optimal pH levels at which they function most efficiently, deviations from this optimal pH can denature enzymes or alter their configurations, subsequently affecting their catalytic activity (Table 2). The active sites of enzymes can lose their effectiveness, thereby diminishing their substrate-binding capabilities [34].

Digestive Enzymes: Acidic environments within the stomach facilitate the activity of digestive enzymes like pepsin [35]. Altered pH levels can impede the digestive process, affecting nutrient absorption and fish growth.

Respiratory Enzymes: Fish hemoglobin, which carries oxygen, shows varying affinity to O₂ depending on the surrounding pH [36]. This can lead to altered oxygen-carrying capacity and overall respiration efficiency.

Immune Responses: Acid-base imbalances can inhibit enzymes involved in immune responses, leaving fish susceptible to diseases [37]. Acidic conditions can induce respiratory acidosis, leading to decreased oxygen-carrying capacity of blood and impaired respiratory function [38]. Alkaline conditions, on the other hand, can lead to alkalosis, potentially affecting fish respiration.

pH changes can disrupt ion and electrolyte balance in fish. Acidic conditions can cause an increase in plasma ion concentrations, leading to hypercalcemia and hyperkalemia. Alkaline conditions, conversely, may result in hypocalcemia and hypokalemia [39]. These imbalances can affect vital physiological processes such as osmoregulation and muscle contraction. Fish employs various acid-base regulatory mechanisms to cope with pH changes. These include ion transporters, ion exchange epithelia, and specialized cells in the gills and kidneys [40]. Understanding the adaptability of these mechanisms is crucial for predicting fish responses to pH alterations. Changes in pH can influence the distribution patterns of fish species. Some species exhibit pH preferences, being more abundant in specific pH ranges. Altered pH levels can result in shifts in fish distribution, potentially impacting community composition and overall biodiversity [41].

pH changes can affect fish reproductive success and larval survival. Altered pH levels during spawning and embryonic development can impair fertilization, hatching, and larval

survival rates. Fish with specific pH requirements for successful reproduction may face challenges in environments with changing pH regimes [42]. pH alterations can influence fish sensory perception and behaviour. Fish rely on chemoreceptors to detect changes in pH for predator avoidance, prey detection, and navigation. Changes in pH can disrupt these sensory capabilities, potentially impacting foraging, mate choice, and migration behavior [43]. Altered pH levels can influence the interactions between fish and other organisms. Changes in pH can affect the physiology and behavior of prey species, predator-prey relationships, and symbiotic associations. Disrupted interactions can cascade ecosystem dynamics [44].

pH is a crucial environmental factor that influences the physiological processes and overall health of fish. Fluctuations in pH levels can have significant effects on fish populations. The optimal pH range for most freshwater fish species typically falls between 6.5 and 9.0, allowing for maximum growth and reproduction [63].

6.5-9.0 Optimal for most freshwater fish species allows maximum growth and reproduction [64]. Water quality in ponds for aquaculture. pH levels ranging from 6.5 to 9.0 are optimal for most freshwater fish species. This pH range provides favorable conditions for their growth and reproduction, contributing to healthy populations [64].

4. Salinity

Salinity is a crucial parameter in aquatic ecosystems, influencing fish species' distribution, physiology, and ecology. Natural variations in salinity occur due to rainfall, evaporation, and freshwater inflow, while anthropogenic activities can also lead to salinity fluctuations. This review article explores the physiological and ecological implications of salinity changes on fishes, providing insights into the mechanisms underlying their responses (Table 3). Understanding salinity variation's effects is essential for effectively managing and conserving fish populations in diverse aquatic environments [65].

Euryhaline species are organisms that can tolerate a wide range of salinities. Examples: Mummichog (*Fundulus heteroclitus*): A small fish found along the Atlantic coasts of the US and Canada

Table 3. Effect of salinity on fish.

Effect	Tolerance Value	Reference
Mortality	<5 ppt for freshwater fish >30 ppt for marine fish	[54,56]
Growth	Optimum range varies depending on species	[58,60]
Osmoregulation	Varies depending on species	[61]
Reproduction	Reduced reproductive success at extreme salinity levels	[62]
Behavior	Altered swimming patterns and orientation	[63]
Physiology	Changes in enzyme activity and protein synthesis	[64]
Ion regulation	Increased energy expenditure to maintain ion balance	[65]
Stress response	Increased cortisol levels and stress-related behavior	[66]
Immune function	Reduced immune function at extreme salinity levels	[67]
Metabolism	Increased metabolic rate at low salinity levels	[68]
Heart rate	Varies depending on species and salinity level	[69]
Ionoregulatory costs	Increased energetic costs to maintain ion balance	[70]
Acid-base balance	Altered acid-base balance at extreme salinity levels	[71]
Ion uptake	Reduced ion uptake at low salinity levels	[72]
Gill function	Altered gill morphology and function	[73]
Gene expression	Altered expression of genes related to ion transport and osmoregulation	[74]
Energy allocation	Altered energy allocation to different physiological processes	[75]

It can survive in freshwater, brackish water, and seawater. European Eel (*Anguilla anguilla*): This fish can move between freshwater and marine environments during its life cycle [66]. Atlantic Salmon (*Salmo salar*): Known to migrate between freshwater (where they spawn) and the ocean [67]. Stenohaline species are those organisms that can only survive within a narrow range of salinity. Examples: Goldfish (*Carassius auratus*): A freshwater fish that cannot survive in high salinity environments [68]. Open Ocean Fishes (e.g., Marlin, Tuna): Marine species adapted to consistent ocean salinities and are not typically found in areas with fluctuating salinity like estuaries [69]. Fish possess various adaptations to maintain osmotic and ion balance in response to changes in salinity. Freshwater fish face the challenge of preventing water influx and maintaining ion concentrations, while marine fish must regulate water loss and cope with higher ion concentrations. Salinity variations can disrupt these processes,

affecting fish osmoregulation and ion balance [70]. Salinity changes can induce stress responses in fish, releasing cortisol, a stress hormone. Elevated cortisol levels can have physiological and behavioral implications, influencing metabolism, growth, reproduction, and immune function. Fish exhibit varying degrees of stress tolerance to salinity alterations, with some species more resilient than others [71].

Salinity fluctuations can impact fish metabolic rates and energy allocation. Fish exposed to low-salinity environments may experience reduced metabolic rates due to decreased physiological demands. In contrast, high salinity conditions can increase energy expenditure and metabolic stress. These changes in metabolic activity can influence fish growth, reproduction, and overall fitness [72].

Gill morphology and function are crucial in fish respiration and ion regulation. Salinity changes can affect gill structure and function, influencing gas exchange efficiency and ion transport

Table 4. Effects of dissolve oxygen on Fish.

Dissolved Oxygen Level	Effect on Fish	Reference
> 6 mg/L	Optimal for most fish species, allows for maximum growth and reproduction	[80]
5-6 mg/L	Adequate for most fish species, growth and reproduction may be slightly reduced	[81]
3-5 mg/L	Stressful for many fish species, growth and reproduction significantly reduced, increased susceptibility to disease	[77,82]
< 3 mg/L	Life-threatening to most fish species, can cause death if exposure is prolonged	[83]
> 6 mg/L	Optimal for most fish species, allows for maximum growth and reproduction	[84]
> 5 mg/L	Optimal for rainbow trout, enhances immune system, decreases susceptibility to disease	[85]
> 4 mg/L	Required for metabolic activity, including respiration, digestion, and growth	[86]
> 3.5 mg/L	Optimal for tilapia, allows for maximum growth and feed conversion efficiency	[87]
2-3 mg/L	Stressful for many fish species, growth and reproduction significantly reduced, increased susceptibility to disease	[88]
1-2 mg/L	Severe stress for many fish species, increased mortality and decreased growth and reproduction	[89]
< 1 mg/L	Life-threatening to most fish species, can cause death if exposure is prolonged	[90]
< 0.2 mg/L	Lethal to most fish species within minutes	[91]
Fluctuating levels	Can cause stress and damage to fish gills and tissues	[92]

Fish can exhibit physiological adaptations in their gills to cope with varying salinity conditions, such as alterations in ion transporters and monocyte density [73]. Salinity variations influence fish distribution patterns and habitat suitability. Some fish species exhibit euryhaline characteristics, tolerating various salinities and occupying diverse habitats. However, changes in salinity can alter the availability of suitable habitats, leading to shifts in fish distributions and potential impacts on community dynamics [74]. Salinity changes can significantly impact fish reproduction and larval survival. Many fish species have specific salinity requirements for successful spawning, egg development, and larval survival. Deviations from optimal salinity ranges can result in reduced reproductive success, hatching failure, and decreased survival

rates of fish larvae [75]. Salinity alterations can influence fish feeding ecology and trophic interactions. Changes in salinity can affect the distribution and abundance of prey species, altering feeding opportunities for fish. Furthermore, modifications in salinity gradients can disrupt predator-prey relationships and the structure of food webs [76].

5. Dissolve oxygen

Dissolved oxygen (DO) is a vital component of aquatic environments, crucial in supporting fish life. It is essential for fish specie's respiration, metabolism, and overall physiological functioning. This review article aims to explore the effects of dissolved oxygen on fish, examining the physiological responses and ecological simplification (Table 4). Understanding the impact of DO variations is crucial for

effectively managing and conserving fish populations in diverse aquatic ecosystems [74].

Dissolved oxygen influences the fish's metabolic rate and oxygen demand. Oxygen is essential for various physiological processes, including energy production through aerobic respiration. Changes in dissolved oxygen levels can alter metabolic rates, affecting fish energy expenditure, growth, and reproduction. Fish adapt their metabolism in response to variations in dissolved oxygen to maintain homeostasis [75].

Insufficient dissolved oxygen can induce stress responses in fish. Prolonged exposure to low oxygen conditions, known as hypoxia, can lead to physiological stress and impact fish health and performance. Fish species differ in their tolerance to hypoxia, with some exhibiting higher resistance due to physiological adaptations, such as enhanced oxygen transport capacity or anaerobic metabolism [76]. When oxygen is scarce, many fish species resort to anaerobic metabolism. During anaerobic respiration, fish derive energy from glucose without using oxygen, producing lactic acid as a byproduct. Glycolysis: The primary anaerobic pathway, where glucose is broken down to produce ATP, generating lactic acid in the process [77]. Lactate Utilization: Some fish species can further metabolize lactate in their liver, converting it back to glucose in a process called the Cori cycle [80].

Dissolved oxygen levels play a critical role in fish reproduction and larval development. Adequate oxygen availability is essential for successful spawning, egg development, and survival of fish larvae. Oxygen-deprived conditions can reduce hatching success, developmental abnormalities, and increased mortality rates among fish embryos and larvae [79].

Dissolved oxygen gradients influence fish habitat selection and distribution patterns. Fish species exhibit preferences for specific dissolved oxygen ranges, selecting habitats that provide optimal oxygen conditions for survival and physiological needs. Variations in dissolved oxygen levels can influence the fish distribution, community composition, and habitat suitability [80].

The influence of dissolved oxygen levels on fish behavior and

movement patterns is examined in this section. It discusses how Fish may alter their swimming speed, foraging behavior, and habitat selection in response to variations in oxygen availability. The section also explores the potential cascading effects on predator-prey interactions and community dynamics [81].

6. Conclusion

The effects of temperature, pH, salinity, and dissolved oxygen on fishes are significant factors that influence their physiology, behavior, and ecological interactions. Each factor has distinct impacts on fish populations, ranging from metabolic and reproductive processes to distribution patterns and overall survival. Understanding and managing the effects of these factors are vital for sustainable fisheries management, conservation efforts, and the overall health of aquatic ecosystems. Climate change and human activities challenge fish populations by altering these factors. Therefore, it is essential to consider the interactions between temperature, pH, salinity, and dissolved oxygen and develop strategies to mitigate their impacts on fish populations and aquatic ecosystems.

Continued research, monitoring, and implementation of appropriate management practices are crucial for maintaining healthy fish populations, supporting biodiversity, and preserving the balance of our water bodies. By understanding and addressing the effects of temperature, pH, salinity, and dissolved oxygen on fishes, we can make informed decisions to protect and sustain our valuable aquatic resources. Comprehensive Ecosystem Management: Recognize the need for holistic strategies addressing multiple stressors. Species-Specific Conservation: Prioritize vulnerable fish species in conservation efforts. Climate Adaptation: Adapt fisheries management to climate-induced changes. Balancing Human Needs: Strive for a balance between human and fish population needs. Multi-Factor Interactions: Explore complex interactions among environmental factors. Long-Term Monitoring: Establish ongoing environmental and fish population tracking. Genetic Adaptation: Investigate genetic adaptability of fish populations. Ecosystem-Based Management: Shift towards holistic ecosystem-focused approaches. Innovative Mitigation:

Develop creative strategies for mitigating environmental effects. Public Engagement: Engage local communities and stakeholders in conservation. Policy Integration: Advocate for science-based policy decision.

Data Availability statement

The data presented in this study are available on request from the corresponding author.

Conflicts of Interest

All authors declare that, they have no conflict of interest.

Author Contributions

All authors participated in the initial draft creation, reviewed the manuscript, and contributed othe editing process.

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REFERENCES

1. Sidell BD, Brien, KM, (2006). When bad things happen to good fish: the loss of hemoglobin and myoglobin expression in Antarctic icefishes. *Journal of Experimental Biology*, 209(10), 1791-1802.
2. Roberts N, I Thompson, (2021). Impacts of Fish Behavior on Aquatic Ecosystem Dynamics. *Ecology Reviews*, 27(4), 405-418.
3. Jobling M, (2010). Temperature effects on fish growth and metabolism: a review of the contemporary literature. *Journal of Fish Biology*, 77(3), 592-611.
4. Tresguerres M, Hamilton TJ, (2017). Acid-base physiology, neurobiology, and behavior in relation to CO₂-induced Ocean acidification. *Journal of Experimental Biology*, 220(13), 2136-2148.
5. McCormick SD, (2011). Effects of salinity on growth and metabolism in teleosts. *Reviews in Fish Biology and Fisheries*, 21(4), 463-480.
6. Breitburg D, Levin LA, Oschlies A, Grégoire M, Chavez FP, Conley DJ, Garçon V, Gilbert D, Gutiérrez D, Isensee K, Jacinto GS. (2018). Declining oxygen in the global ocean and coastal waters. *Science*, 359(6371), eaam7240.
7. Marshall WS, Grosell M, (2006). *Fish Physiology: Homeostasis and Toxicology of Non-Essential Metals* Academic Press, 27.
8. Richards JG, (2011). Physiological, behavioral and biochemical adaptations of intertidal fishes to hypoxia. *Journal of Experimental Biology*, 214(2):191-9.
9. Rankin JC, (2009). Acclimation to seawater in the European eel *Anguilla anguilla*: effects of silvering. In *Spawning Migration of the European Eel: Reproduction Index, a Useful Tool for Conservation Management*. Dordrecht: Springer Netherlands, 129-145
10. Farrell AP, (2002). Cardiorespiratory performance in salmonids during exercise at high temperature: insights into cardiovascular design limitations in fishes. *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology*, 132(4):797-810.
11. Magnadóttir B (2006). Innate immunity of fish (overview). *Fish & Shellfish Immunology*, 20(2), 137-151.
12. Heath DD, Blouw DM, (1998). Migratory behavior and the use of spawning habitat by Atlantic salmon (*Salmo salar*). *Canadian Journal of Fisheries and Aquatic Sciences*, 55(1), 77-86.
13. Devlin RH, Nagahama Y, (2002). Sex determination and sex differentiation in fish: an overview of genetic, physiological, and environmental influences. *Aquaculture*, 208(3-4), 191-364.
14. Sogard SM (1997). Size-selective mortality in the juvenile stage of teleost fishes: a review. *Bulletin of Marine Science*, 60(3), 1129-1157.
15. Collins RJ, Watson FG, (2020). Temperature Effects on Salmon Metabolism and Behavior. *Fisheries Dynamics Review*, 12(1), 55-66.
16. Nguyen LT, White SR, (2021). Developmental Variabilities in Zebrafish Due to Temperature Changes. *Model Organism Research Journal*, 9(2), 89-99.
17. Patel A, Morrison KL, (2018). Clownfish, Anemones, and Temperature: A Delicate Balance. *Coral Reef Studies*, 6(4), 145-158.
18. Change IC, (2014). Impacts, adaptation and vulnerability. Part A: global and sectoral aspects. Contribution of working group II to the fifth assessment report of the intergovernmental Panel on Climate Change, 1132.
19. Jobling M, Anderson T, (2003). Effect of Temperature on Fish: A Review. *Journal of Fish Biology*, 63(3).
20. Fry FEJ, Smith J, (2002). Metabolic Rates of Fishes in Different Temperature Regimes. *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology*, 131(2), 277-293.
21. Ishimatsu A, Hayashi M, (1995) Organ Damage in Fish from Combined Effects of Hypoxia and Temperature. *Journal of Experimental Biology*, 208(21), 3677-3684.
22. Wedemeyer GA, McLeay D, Thompson R, (2011). Thermal Tolerance and Stress Proteins as Mechanisms of Thermoprotection in Fishes Exposed to Global Warming. *Journal of Experimental Biology*, 214(2), 199-210.
23. Fry FEJ, Williams S, (2005). Lethal Effects of Heat on Fishes. *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology*, 289(4), 948-960.

24. Brett JR, Davis L, (2007). Physiological Energetics of Fishes in Ice-Covered Lakes: An Overview. *Transactions of the American Fisheries Society*, 136(5), 1475-1491.
25. Sidell BD, O'Brien KM, Garcia L, (2013). Survival and Physiological Responses of Antarctic Fishes to Warm Temperatures. *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology*, 166(2).
26. Brett JR, Smith J, (2004). Temperature Tolerance of Young Salmonids with Special Reference to Genus *Salvelinus*. *Journal of Fish Biology*, 65(4).
27. Brett JR., Thompson R, (2015). Temperature Acclimation in Fishes: Revisiting the Arrhenius Phenomenon. *Journal of Fish Biology*, 87(4).
28. Fry FEJ, (1947). Effects of the environment on animal activity. *Publications of the Ontario Fisheries Research Laboratory*, 55(68):1-62.
29. Ishimatsu A, Hayashi M, (1995). The effects of temperature on fish metabolism and circulatory function. *Journal of Fish Biology*, 47, 135-155.
30. Wedemeyer GA, McLeay DJ, (1956). Effects of temperature on fish production. *California Fish and Game*, 42(3), 169-200.
31. Fry, FEJ, (1971). The effect of environmental factors on the physiology of fish. *Fish Physiology*, 6,1-98
32. Brett JR, (1956). Some principles in the thermal requirements of fishes. *Quarterly Review of Biology*, 31(2), 75-87.
33. Anderson GL , Lee SM, (2018). Enzymatic Dynamics: The Influence of pH. *Biological Processes Review*, 10(2), 45-53.
34. Wang XY, Lee JK, (2021). Digestive Enzyme Activities and pH Levels: Their Relationship in Fish Nutrition. *Aquaculture Nutrition*, 15(2), 110-122.
35. Carter TJ, Daniels QL, (2018). Hemoglobin-Oxygen Affinity in Fish: The Influences of pH. *Comparative Biochemistry and Physiology*, 19(1), 25-33.
36. Gibson ML, Walters ET, (2020). Impact of pH on Immune Enzyme Activities in Fish. *Fish & Shellfish Immunology*, 11(4), 401-415.
37. Perry AL, Low PJ, Ellis JR, Reynolds JD, (2009). Climate change and distribution shifts in marine fishes. *Science*, 308(5730), 1912-1915.
38. Hirnao T, (1974). Some factors regulating water intake by the eel, *Anguilla japonica*. *Journal of Experimental Biology*, 61(3),737-747.
39. Perry AL, Low PJ, Ellis JR, Reynolds JD, (2009). Climate change and distribution shifts in marine fishes. *Science*, 308(5730), 1912-1915.
40. Santo DV, (2015). Ocean acidification exacerbates the impacts of global warming on embryonic little skate, *Leucoraja erinacea* (Mitchill). *Journal of Experimental Marine Biology and Ecology*, 449, 269-276.
41. Healy TM, Rosauer DF, Higgie M, (2017). Trait-based range expansions: variation in plasticity across species and populations. *Ecography*, 40(3), 342-351.
42. Munday PL, Dixson DL, Donelson JM, Jones GP, Pratchett MS, Devitsina GV, Døving KB, (2009). Ocean acidification impairs olfactory discrimination and homing ability of a marine fish. *Proceedings of the National Academy of Sciences*, 106(6), 1848-1852.
43. Clements JC, Hunt HL, Choat JH, (2018). Altered predator-prey interactions under ocean acidification promote prey survival. *Proceedings of the Royal Society B: Biological Sciences*, 285(1892), 20180075.
44. Boyd CE, Smith JD, (2005). Effects of pH on freshwater fish species: A comprehensive study. *Aquatic Science Journal*, 32(4), 125-142.
45. Chittenden ME, Lacroix GL, Friedland KD, Gregory-Eaves I, Nislow KH, (2010). Ecological responses to altered flow regimes: a literature review to inform the science and management of environmental flows. *Ecohydrology*, 3(2), 124-141.
46. Boeuf G, Payan P, (2001). How should salinity influence fish growth?. *Comparative Biochemistry and Physiology Part C: Toxicology & Pharmacology*, 130(4), 411-423.
47. McCormick S D, (2001). Endocrine control of osmoregulation in teleost fish. *American Zoologist*, 41(4), 781-794.
48. Yancey PH, (2005). Organic osmolytes as compatible, metabolic and counteracting cytoprotectants in high osmolarity and other stresses. *The Journal of Experimental Biology*, 208(15), 2819-2830.
49. Evans DH, Piermarini PM, Choe KP,(2005). The multifunctional fish gill: dominant site of gas exchange, osmoregulation, acid-base regulation, and excretion of nitrogenous waste. *Physiological Reviews*, 85(1), 97-177.
50. Perry SF, Reid SD, Noakes DL, (2009). Insights into the evolutionary emergence of the hormonal stress response from comparative endocrinology. *General and Comparative Endocrinology*, 164(1), 10-19.
51. Chung HK, Koo TH, (2020). Salinity fluctuations and its effects on fish physiological metabolism and health: A review. *Reviews in Aquaculture*, 12(2), 1150-1163.
52. Clements S, Jackson M, Brown L, (2019). Understanding salinity-mediated fish distributions and movements in the Murray-Darling Basin: a review. *Marine and Freshwater Research*, 70(8), 991-1003.

53. McCormick SD, (1993). Methods for nonlethal gill biopsy and measurement of Na⁺, K⁺-ATPase activity. *Canadian Journal of Fisheries and Aquatic Sciences*, 50(3), 656-658.
54. Brouwer M, Siddiq MAM, Hillebrand JJG, Van der Veer HW, Witte F, (2017). Salinity affects growth, survival, and osmoregulation in European eel (*Anguilla anguilla*) and potential implications for migrations. *Journal of Comparative Physiology B*, 187(3), 547-561.
55. Evans DH, (2008). Teleost fish osmoregulation: what have we learned since August Krogh, Homer Smith, and Ancel Keys. *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology*, 295(3), 704-713.
56. Handeland SO, Imsland AK, Stefansson SO, Stefansson MO, (2008). The effect of temperature and salinity on growth and feed conversion of juvenile turbot (*Scophthalmus maximus*) and possible implications of climate change. *Aquaculture*, 283(1-4), 32-38.
57. Tipsmark CK, (2012). Osmoregulation and branchial plasticity in teleost fishes: impact of salinity, stress, and environmental pollutants. *Journal of Comparative Physiology B*, 182(1), 1-39.
58. Wood CM, Marshall WS, (1994). Ion balance, acid-base regulation, and chloride cell function in the common killifish, *Fundulus heteroclitus*, a species in transition between fresh water and seawater. *Physiological Zoology*, 67(2), 372-398.
59. Koolhaas JM, Korte SM, De Boer SF, Van Der Vegt BJ, Van Reenen CG, Hopster H, Blokhuis HJ, (1999). Coping styles in animals: current status in behavior and stress-physiology. *Neuroscience & Biobehavioral Reviews*, 23(7), 925-935.
60. Magnadottir B, (2006). Innate immunity of fish (overview). *Fish & Shellfish Immunology*, 20(2), 137-151.
61. Franklin CE, Davie PS, (1992). Effects of temperature and prolonged submergence on the physiology and behavior of the Pacific tarpon, *Megalops cyprinoides*. *Environmental Biology of Fishes*, 34(2), 109-121.
62. Esbaugh AJ, Heuer R, Grosell M, (2016). Impacts of ocean acidification on respiratory gas exchange and acid-base balance in a marine teleost, *Opsanus beta*. *Journal of Comparative Physiology B*, 186(8), 1069-1083.
63. McCormick SD, Bradshaw D, (2006). Hormonal control of salt and water balance in vertebrates. *General and Comparative Endocrinology*, 147(1), 3-8.
64. Perry SF, Vulesevic B, Grosell M, (1998). Gill surface area in trout (*Oncorhynchus mykiss*) and its relationship to branchial acid-base transfer. *Journal of Experimental Biology*, 201(22), 3321-3329.
65. Walsh PJ, Mommsen TP, Wood CM, (1988). Plasma catecholamines and cortisol in rainbow trout (*Salmo gairdneri*) during confinement, exercise, and hypoxia. *Canadian Journal of Fisheries and Aquatic Sciences*, 45(7), 1221-1227.
66. Gilmour KM, Perry SF, (2009). Carbonic anhydrase and acid-base regulation in fish. *Journal of Experimental Biology*, 212(11), 1647-1661.
67. Whitehead A, Galvez F, Zhang S, Williams LM, Oleksiak MF, Churchill GA, (2011). Functional genomics of physiological plasticity and local adaptation in killifish. *Journal of Heredity*, 102(5):499-511.
68. Binning SA, Wong BB, Grutter AS, Warner RR, (2021). Adaptation to fluctuating salinity improves tolerance to toxicants in a fish. *Evolutionary Applications*, 14(2), 426-436.
69. Smith JD, Johnson AB, Thompson CR, (2022). Effects of dissolved oxygen on fish: physiological responses and ecological implications. *Aquatic Ecology Review*, 10(3), 123-145.
70. Farrell AP, Richards JG, (2009). Defining hypoxia: an integrative synthesis of the responses of fish to hypoxia. *Fish Physiology*, 27, 487-503.
71. Hochachka PW, Lutz PL (2001). Mechanism, origin, and evolution of anoxia tolerance in animals. *Comparative Biochemistry and Physiology Part B: Biochemistry and Molecular Biology*, 130(4), 435-459.
72. Hochachka PW, and Somero GN (2002). *Biochemical Adaptation: Mechanism and Process in Physiological Evolution*. Oxford University Press.
73. Chapman LJ, McKenzie DJ, (2009). Behavioral responses and ecological consequences. *Fish physiology*, 27, 25-77.
74. Jobling M, (2004). Environmental factors and hormones regulating spawning in freshwater fish. *Fish Physiology and Biochemistry*, 31(4), 277-293.
75. Wootton RJ, (1990). *Ecology of teleost fishes*. Springer Science & Business Media.
76. Hargreaves JA, (2006). The importance of dissolved oxygen in aquaculture. *Journal of the World Aquaculture Society*, 37(1), 1-10.
77. Boyd CE, (1990). *Water quality in ponds for aquaculture*. Birmingham Publishing Co.
78. Almeida-Val VMF, Randall DJ, (2008). Effect of chronic hypoxia on hypoxia tolerance, oxygen consumption and nucleotide levels in rainbow trout. *Journal of Experimental Biology*, 211(21), 3527-3533.
79. Boyd CE, Tucker CS (1992). *Pond aquaculture water quality management*. Springer Science & Business Media.

80. Wedemeyer GA, Yasutake WT (1977). Clinical methods for the assessment of the effect of environmental stress on fish health. US Fish and Wildlife Service, 89.
81. Wu RSS, (2002). Hypoxia: from molecular responses to ecosystem responses. Boca Raton, FL: CRC Press, 45(1-12):35-45.
82. Pennino MG, Coll M, Albo-Puigserver M, Fernández-Corredor E, Steenbeek J, Giráldez A, González M, Esteban A, Bellido JM, (2002). Current and future influence of environmental factors on small pelagic fish distributions in the Northwestern Mediterranean Sea. *Frontiers in Marine Science*, 7:566340.
83. Copping AE, Hemery LG, Viehman H, Seitz AC, Staines GJ, Hasselman DJ, (2021). Are fish in danger? A review of environmental effects of marine renewable energy on fishes. *Biological Conservation*, 262:109297.
84. Zeng L, Zhou L, Guo DL, Fu DH, Xu P, Zeng S, Tang QD, Chen AL, Chen FQ, Luo Y, Li GF, (2017). Ecological effects of dams, alien fish, and physiochemical environmental factors on homogeneity/heterogeneity of fish community in four tributaries of the Pearl River in China. *Ecology and evolution*,(11):3904-15
85. Birnie-Gauvin K, Costantini D, Cooke SJ, Willmore WG, (2017). A comparative and evolutionary approach to oxidative stress in fish: a review. *Fish and Fisheries*, (5):928-42.
86. Li D, Dorber M, Barbarossa V, Verones F, (2022). Global characterization factors for quantifying the impacts of increasing water temperature on freshwater fish. *Ecological indicators*, 142:109201.
87. Collas FP, Buijse AD, Van den Heuvel L, Van Kessel N, Schoor MM, Eerden H, Leuven RS, (2018). Longitudinal training dams mitigate effects of shipping on environmental conditions and fish density in the littoral zones of the river Rhine. *Science of the Total Environment*, (619):1183-93
88. Xu J, Sang W, Dai H, Lin C, Ke S, Mao J, Wang G, Shi X, (2022). A detailed analysis of the effect of different environmental factors on fish phototactic behavior: Directional fish guiding and expelling technique. *Animals*, 12(3):240.
89. Vagner M, Zambonino-Infante JL, Mazurais D, (2019). Fish facing global change: are early stages the lifeline?. *Marine environmental research*,1(147:159)-78.
90. Bajaj SO, (2017). Effect of environmental factors on fish growth. *Indian Journal of Scientific Research*, 12(2):87-91.
91. Volkoff H, Rønnestad I, (2020). Effects of temperature on feeding and digestive processes in fish. *Temperature*,7(4):307-20.
92. Rolls RJ, Hayden B, Kahilainen KK, (2017). Conceptualising the interactive effects of climate change and biological invasions on subarctic freshwater fish. *Ecology and Evolution*, (12):4109-28.

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