

Review article

# ZnO Nanoparticles Impact on Organ Systems in Rats: A Comprehensive Exploration of Diverse Exposure Pathways

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## Abstract

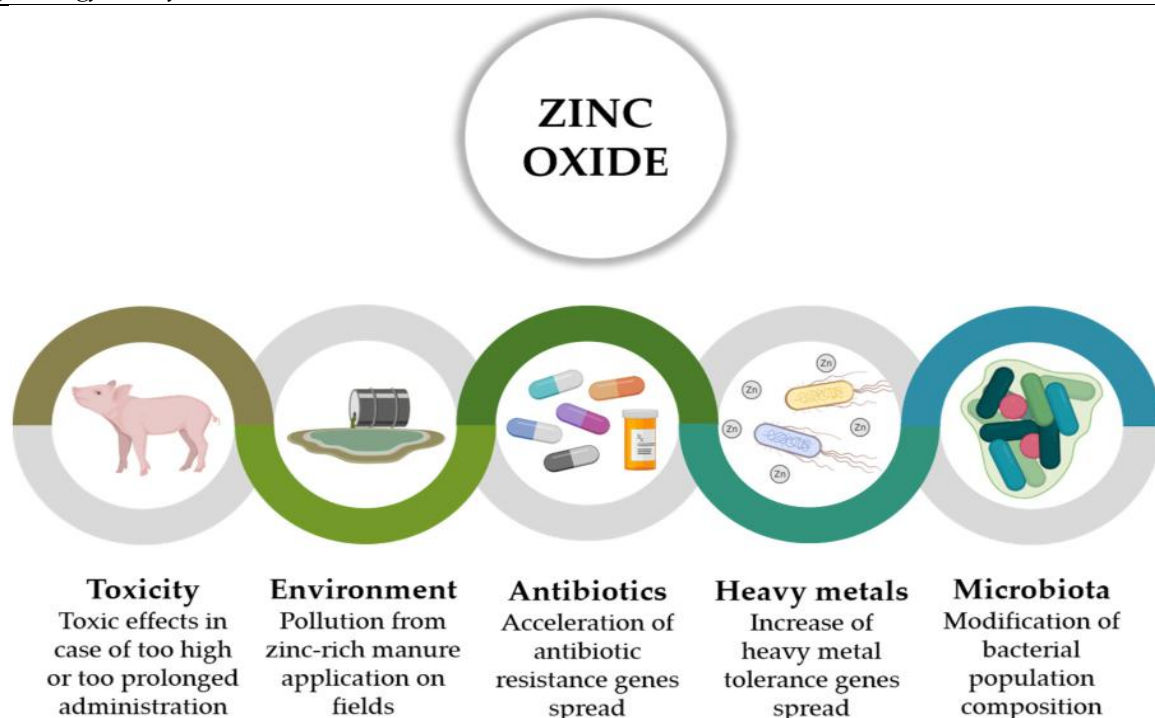
The synthesis and utilization of nanomaterials with precise spatial dimensions on the nanoscale are pivotal in the field of nanotechnology. In recent years, metal oxide nanoparticles have become increasingly common, raising concerns in both scientific community and the general public about their potential harm to the environment and living organisms. Despite this, there are still significant debates and misconceptions regarding the adverse effects and mechanisms of these nanoparticles. To facilitate their safe and responsible use, it is imperative to gain a comprehensive understanding of their adverse effects. This review aims to provide an overview of the biological fate of zinc oxide (ZnO) nanoparticles in rats through various exposure routes, shedding light on their toxicological consequences and the underlying mechanisms of toxicity. Despite the fact that ZnO nanoparticles have a propensity to target organs such as the liver, kidneys, and lungs, it is noteworthy that higher concentrations of zinc are detected in these tissues following exposure via various routes. The liver plays a central role in the metabolism of ZnO nanoparticles. Multiple exposure routes, including oral, intraperitoneal, intravenous, and intratracheal routes, have been shown to induce liver damage, along with adverse effects on the kidneys and lungs when exposure occurs via airways. A significant toxicological mechanism associated with ZnO nanoparticles involves the generation of reactive oxygen species (ROS) and the subsequent initiation of oxidative stress. ROS production can result from both excessive release of Zn<sup>+2</sup> ions and the particulate effect stemming from the semiconductor or electronic properties of ZnO nanoparticles. The potential for surface coatings and modifications holds the promise of further expanding the range of biomedical applications for ZnO nanoparticles, opening up exciting possibilities for futuristic medical treatments, including targeted drug delivery, advanced imaging techniques, and diagnostics.

**Keywords:** Albino rats, intravenous route, airway exposure, potential infection, hepatic toxicity.

## 1. Introduction

Nanotechnology is a term used to define areas of science and engineering in which phenomena occurring at nanoscale dimensions are used in the design, characterization, manufacture, and applications of materials, structures, devices, and systems [1]. It has the potential to revolutionize the medical research industry and open up new fields for the betterment of humanity [1,2,3]. The three main categories of

nanotechnology are nanotools, nanodevices, and nanostructured materials. Nanotechnology relies on a wide range of techniques, such as computer modeling, surface science, supramolecular chemistry, nanolithography, synthetic approach, and analytical tools. Nanoelectronics, nanospintronics, nanosensors, nano-optical electronics, and nano-drug delivery systems are all examples of nanodevices.



**Figure 1.** Risks associated with pharmaceutical ZnO utilization [4].

Some examples of nanostructured materials are nanowires, nanoparticles, fullerene, carbon nanotubes, graphene, nanocomposites, thin solid films, nano-patterned surfaces, and supramolecular systems [5]. Different morphologies of ZnO nanoparticles, such as nanoflake, nanoflower, nanobelt, nanorod, and nanowire, have been reported [6, 7, 8].

ZnO is a white inorganic substance which is soluble in acidic or alkaline solutions but insoluble in water. It doesn't naturally occur in large amounts [9]. There are numerous ways to make ZnO nanoparticles, including thermal evaporation, gas evaporation, hydrothermal, the vapor-liquid-solid process, self-combustion, simple thermal sublimation, and green synthesis [10]. Moreover, the simplicity, cost-effectiveness, and eco-friendly nature of green synthesizing ZnO nanoparticle synthesis using extracts from different plants and fruits have garnered significantly attention and interest [11]. Several issues have been highlighted by the extensive and extended use of ZnO at pharmaceutical levels in pig husbandry, including ZnO nanoparticle's advantages are lost with excessive or prolonged ZnO intake, which also increases the risk of harmful consequences [12, 13]. Due to

the excessive Zn buildup in animal organs such the pancreas, liver, and kidney [14], which is susceptible to Zn excess [13]. ZnO nanoparticles their increasing application has prompted safety concerns [15]. Size, surface properties, solubility, and mode of exposure are the most important factors in determining the toxicity of metal oxide nanoparticles. The biological fate and toxicity of ZnO nanoparticles upon exposure via various mechanisms must be understood. Workers are most likely to be exposed to ZnO nanoparticles since they are used as food additives and packaging materials ingestion, in addition to the more common routes of exposure (inhalation and skin contact) [16]. If you consume ZnO nanoparticles orally, they will dissolve in your stomach's acidic environment (pH 1.5-2.0) and  $Zn^{+2}$  will be absorbed into your blood [17]. ZnO nanoparticles are likely absorbed in both ionic and particle forms after oral administration to rats [18]. After being injected intraperitoneally, ZnO nanoparticles are taken up by the body as ions rather than particles. After oral and intraperitoneal administration,  $Zn^{+2}$  is taken up by the liver via the first-pass effect and subsequently redistributed. By dissolving in the acidic lining fluid of the lungs, ZnO

nanoparticles are able to cross the alveolar membrane and enter the circulation after being inhaled [19, 20]. Inhaled ZnO nanoparticles pass the alveolar barrier and enter the circulation because they dissolve in the acidic lining fluid of the lungs. Skin has a pH that varies from the surface to the stratum corneum, which may lead to the dissolution of topically administered ZnO nanoparticles and the dermal absorption of Zn [21].

ZnO nanoparticles are particularly useful in biomedical applications due to their unique properties, such as their size similarity to biomolecules, availability of functionality over wide surfaces, and quantum size impact. ZnO quantum dots can be used for medicine delivery and bioimaging thanks to their high biocompatibility, low toxicity, and good stability [22]. The potential health benefits of ZnO nanoparticles as an antibiotic, nutritional supplement, and food additive are also the subject of ongoing study [23] characteristics are present in ZnO Nanoparticles [24, 25]. They are employed to get rid of aquatic vegetation Applications like medication delivery, cancer prevention, diabetes prevention, anti-diabetic, anti-bacterial, and agronomic properties [26]. Although ZnO is employed for targeted drug delivery, its cytotoxicity limitation has not yet been overcome [27]. They exhibit stronger antibacterial effects than chemically produced ZnO nanoparticles [28, 29, 30]. Additionally, they have been used in the production of rubber, paint, water purification, protein adsorption, and dentistry applications. piezoelectric and pyroelectric that is immune to all types of eradication methods, including physical, chemical, and mechanical ones [31].

## **2. Different Exposure Routes**

Emerging pollutants having ecological and toxicological consequences on individuals, communities, and diverse ecosystems, manufactured nanoparticles, and more especially metal oxide nanoparticles, are finding ever-increasing uses in industrial and consumer products [32, 33].

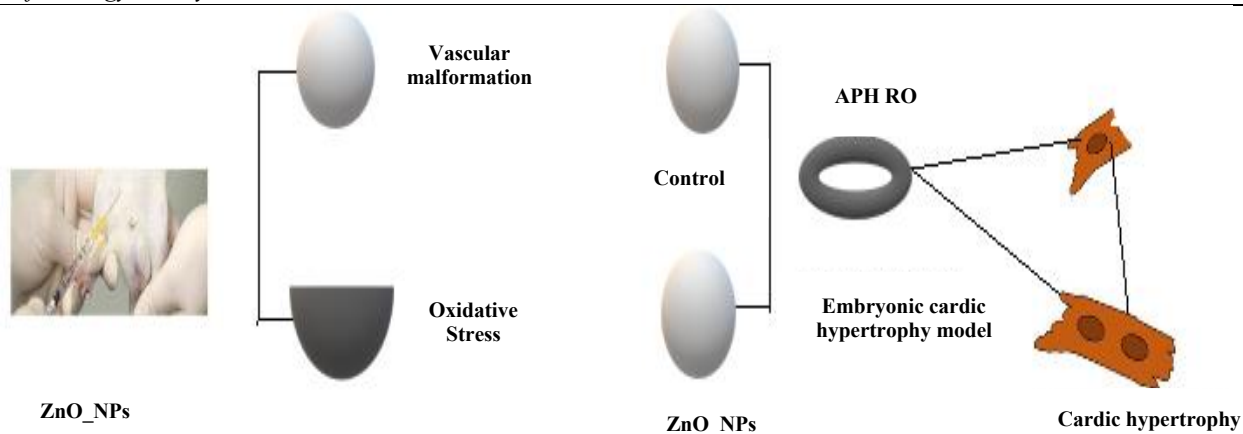
### **2.1 Intraperitoneal Administrations**

According to Li et al. [34] mice were injected intraperitoneally with ZnO nanoparticles (average size 93 nm)

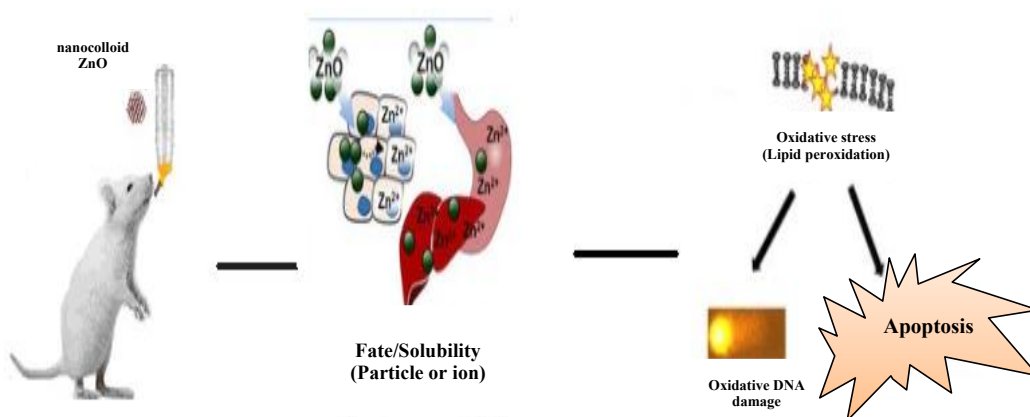
at a dose of 2.5 g/kg, and the findings revealed that Zn accumulated in all organs except the brain (blood-brain barrier). Zn concentrations were highest in the liver, then in the spleen, the lungs, the kidneys, and finally the heart cardiac vascular dysfunction as shown in Figure (2). According to Lin et al. [35] ZnO nanoparticles (size 47.8 nm, dosage 10 mg kg<sup>-1</sup>) accumulated in the liver, lung, kidneys, spleen, and heart 6 hours after a single intraperitoneal injection. Amara et al, studied the effect of intraperitoneal injection of ZnO nanoparticles (size 20-30 nm, dosage 25 mg kg<sup>-1</sup>) and they were found no accumulation of Zn in the liver or kidneys of the rats [36]. Elshama et al. [36] found that long-term intraperitoneal injection of ZnO nanoparticles generated histological and ultrastructural abnormalities in the brains and spinal cords of rats, with the severity of these changes dependent on the dosage and the generation of reactive oxygen species [37].

### **2.2 Oral Administrations**

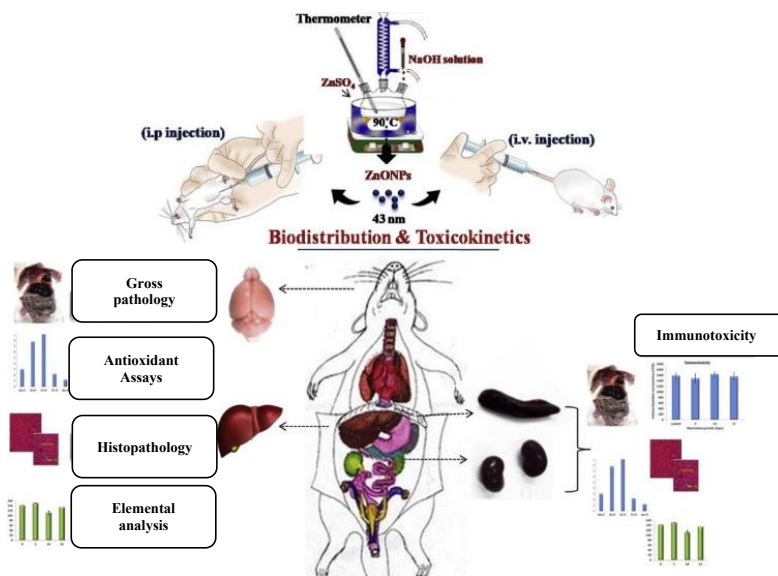
Baek et al. [18] studied the ZnO Nanoparticles (20 and 70 nm) accumulated in the kidneys, liver, and lungs of rats after a single oral treatment Zn levels in all these organs were significantly increased 6-24 hours after a low dosage (50 and 300 mg kg<sup>-1</sup>) of ZnO nanoparticles. After two days of receiving a substantial dosage (2000 mg kg<sup>-1</sup>), considerable buildup occurred in the liver and kidneys; however, by day seven, levels had returned to normal (about 20 g/g in the liver and around 10 g/g in the kidneys). Earlier research has demonstrated that, upon acute oral exposure, nano-forms of certain particles are more hazardous than their micro-counterparts. Liu N et al. [17] found the level of Zn content accumulation in the heart, liver, spleen, lungs, kidneys, and brain of exposed mice, after receiving a single oral dosage of 45 mg kg<sup>-1</sup> of ZnO nanoparticles (size 27.54.1 nm), The tissue distribution pattern of ZnO nanoparticles was found to be different from that of ZnCl<sub>2</sub> with a greater concentration in the lungs and a lower concentration in the kidneys and liver which leads to oxidative stress as well as DNA damage shown in Figure (3). ZnO nanoparticles (size 40 nm, dosage 134.2-536.8 mg kg<sup>-1</sup> day<sup>-1</sup>) were distributed to the liver and kidneys in rats



**Figure 2.** Intraperitoneal ZnO nanoparticles induces vascular malformation and oxidative stress.



**Figure 3.** Oral ingestion leading to stomach digestion-induced apoptosis and DNA damage.



**Figure 4.** Intravenous and intraperitoneal injection of ZnO nanoparticles induces acute toxicity in vital organs including liver, lungs, kidneys, and other body systems [38].

given repeated oral doses for 13 weeks, although lung distribution was not mentioned.

After being given ZnO nanoparticles (size 30 nm, dose 300 mg kg<sup>-1</sup>) orally for 14 days in a row, mice saw their liver Zn content significantly increase and their kidney Zn content somewhat increase [16]. According to Choi et al. [15] a single oral dose of either 3 or 30 mg kg<sup>-1</sup> of ZnO nanoparticles showed that these particles were predominantly distributed in the liver, kidneys, and lungs. Following oral administration, it was observed that ZnO nanoparticles had limited absorption in the gastrointestinal tract (GIT) and were primarily excreted in feces. [39].

Pasupuleti et al. [40] considered that ZnO nanoparticles accumulated in the liver of mice after 14 days of oral treatment of 30 nm ZnO nanoparticles, and they also caused oxidative stress. Sprague Dawley (SD) rats were also given oral administration of ZnO nanoparticles for 14 days. They discovered that rats treated with low doses of nanoparticles had higher rates of lesions in the liver, pancreas, heart, and stomach than rats treated with high doses; however, high dosages of the micro-sized nanoparticles produced more lesions than the low one.

### **2.3 Intravenous Administrations**

Lee J et al. [41] were used rats for checking the effects of intravenous injections of 5, 10, and 20 mg kg<sup>-1</sup> of ZnO nanoparticles on dams and fetuses from gestation day 6 to day 20. Twenty dams in the 20 mg kg<sup>-1</sup> treatment group lost two of them while under treatment. In treated dams, hematological analysis and serum biochemistry revealed dose-dependent damage. Tubular dilatation in the kidneys, extremely hemopoiesis in the liver, and multifocal mixed cell infiltration and thrombosis in the lung were all discovered by histopathological study of treated dams.

Yeh et al. [38] when injected intravenously into mice, radioactive ZnO nanoparticles that emit gamma rays were mostly localized in the lungs, with some also found in the organs responsible for digestion and detoxification. The distribution of ZnO nanoparticles at 24 hours after injection

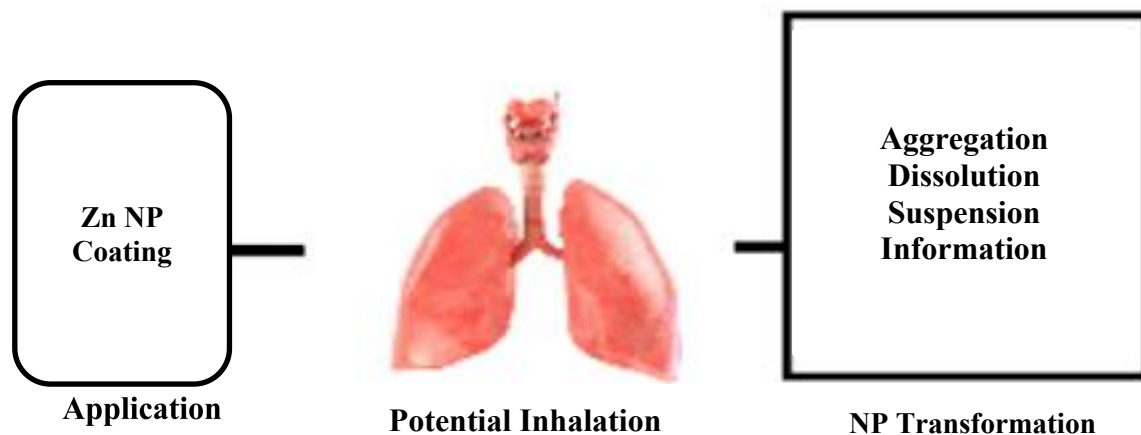
showed the largest amounts in the lungs and liver. ZnO nanoparticles (size 10 and 70 nm, dosage 120 g mouse<sup>-1</sup>) were injected intravenously and studied for their effects on mice over time (days, weeks, and months).

Radioactive ZnO nanoparticles that release gamma rays were predominantly located in the lungs after being injected intravenously into mice, with some also identified in the organs responsible for digesting and detoxification. After being administered intravenously into mice, radioactive ZnO nanoparticles that produce gamma rays were found mostly in the lungs, with some being found in the digestive and detoxifying organs as shown in Figure 4 were given a single intravenous injection, and by the next day, nanoparticles could be found throughout the mice's bodies, including their blood, liver, spleen, lungs, brain, and heart [38].

Fujihara J et al. [42] were administered intravenously ZnO nanoparticles (size 58.5 nm, 0.2 mg kg<sup>-1</sup>) to mice, and their short-term tissue distribution in the lungs, liver, kidneys, and spleen was evaluated for up to 1 hour after administration Liver and lung Zn levels peaked 5 minutes after the treatment, kidney, and spleen Zn levels peaked 15 minutes after the administration, and tissue Zn levels peaked 1 hour after the dose. They also reported that Zn tissue accumulation over time 6 days following intravenous injection of ZnO nanoparticles (0.05 or 0.2 mg kg<sup>-1</sup>). Zn levels were only substantially higher in the kidneys after one day at 0.05 mg kg<sup>-1</sup> compared to the control group. Zn content was considerably elevated in the liver and spleen after just one day and six days at a level of 0.2 mg kg<sup>-1</sup>.

### **2.4 Inhalation Exposure**

Vysloužil et al., [43] were reported that ZnO nanoparticles (size 374.2 nm, dosage 6.46104 and 1.93106 particles/cm<sup>3</sup>) to enter rats organs from ambient air at two different doses, with the lower dose considerably increasing Zn content in the liver and at higher dose significantly increasing Zn content in the lungs. Konduru et al., were injected intratracheally ZnO nanoparticles (size: 4.62.5 nm, dose: 1 mg kg<sup>-1</sup>) into rats and their tissue distribution was mapped out. The transfer of <sup>65</sup>Zn to the skeletal muscle, bone, kidneys, liver, and skin occurred on day 2. It was injected into the skin, bones, and muscles on



**Figure 5.** Inhaled ZnO nanoparticles directly induce lung injury.

days 7 and 28 [44]. Wang et al., were injected intratracheally ZnO nanoparticles (size 4218 nm, dosage  $2.5 \text{ mg kg}^{-1}$ ) in the lungs and liver of mice [19]. Depending on the particle size and solubility, ZnO nanoparticles induced inflammatory and fibrotic responses in the tracheobronchial and alveolar tissues after inhalation. Lung fluid is acidic, so when ZnO nanoparticles were dissolved in it, their concentration increased and they were hazardous to the lungs [45]. Fujihara J., ZnO nanoparticles inhalation experiments revealed negligible lung cytotoxicity, histopathologic alterations, or pulmonary inflammation. ZnO nanoparticles have likely been dissolved in the respiratory system following inhalation if there is a higher Zn content in the BAL fluid and lungs. The toxicity of ZnO nanoparticles was significantly influenced by the exposure concentration, exposure mode, and time post-exposure. To conclude that ZnO nanoparticles had low sub-chronic toxicity via inhalation, exposure for 13 weeks at a cumulative dose of  $10.9 \text{ mg kg}^{-1}$  resulted in increased lung cellularity, but other markers of toxicity did not differ from sham-exposed animals [46].

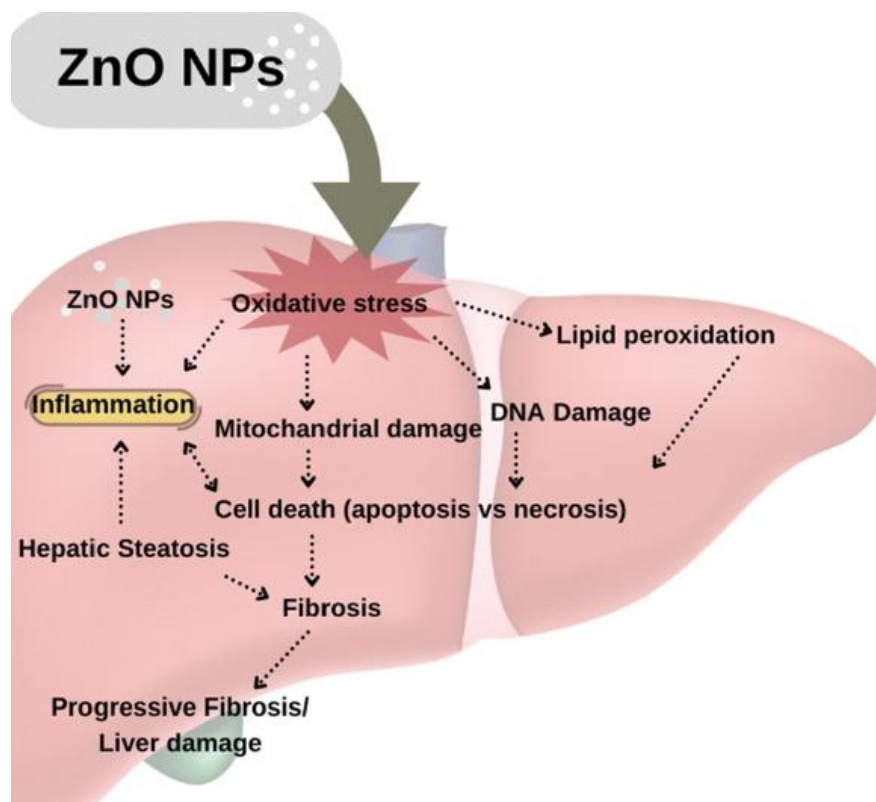
### 3. Toxicological effects ZnO nanoparticles on various organs of Rats

ZnO nanoparticles have been shown to exert harmful effects on different cells, including membrane damage, an inflammatory response, DNA damage, and apoptosis. According to recent research, the release of  $\text{Zn}^{+2}$  ions is what causes ZnO nanoparticles to be poisonous.

#### 3.1 Effects on Body Weight and Organ Weight

Treatment of rats orally with  $536.8 \text{ mg kg}^{-1} \text{ day}^{-1}$  for 13 weeks resulted in a reduction in body weight [47]. Rats (size 12-90 nm) and mice (size 205 nm) were given  $30.3 \text{ mg kg}^{-1}$  intraperitoneally on a daily basis for 28 days, and 1, 10, and  $100 \text{ mg kg}^{-1}$  intraperitoneally daily for 14 days [48]. Rats and mice have been used as examples. After a single oral dosage of  $2000 \text{ mg kg}^{-1}$  (size 20 and 70 nm), rats lost a small amount of weight. At 14 days post-exposure, mice given a single intragastric dose (size 205 nm,  $100 \text{ mg kg}^{-1}$ ) and rats given repeated intraperitoneal and intravenous exposure (size 1290 nm, dose  $30.3 \text{ mg kg}^{-1}$  for 14 and 28 days) showed decreased organ weight of the heart, liver, spleen, lungs, kidneys, and brain [48]. After  $400 \text{ mg kg}^{-1}/\text{day}$  of nanoparticles were administered to the dams, their body weight dramatically fell. They also ate less after receiving 200 and  $400 \text{ mg kg}^{-1} /\text{day}$  of nanoparticles, and after  $400 \text{ mg kg}^{-1} /\text{day}$  of nanoparticles, their liver and adrenal gland weights also rose [49].

Jo et al. [49] were exposed rats to ZnO nanoparticles ( $500 \text{ mg kg}^{-1} \text{ bw}$ ) with a size smaller than 100 nm. Additionally, the zinc concentration in dams and offspring's bodies was assessed. Rats given nano-ZnO treatment had lower pup weights, fewer live births, and higher fetal resorption rates. The liver and kidney of puppies as well as the mammary tissue of mothers were given ZnO nanoparticles. These findings suggest that nanoscale ZnO nanoparticles [50]. Wang et al. [50] found that 50 and  $500 \text{ mg kg}^{-1}$  nano ZnO illustrated increases in body weight while  $5000 \text{ mg kg}^{-1}$  showed declines in body weight, indicating that high dosages of ZnO nanoparticles in the diet



**Figure 6.** Graphical representation of the effects of ZnO nanoparticles on the liver [51].

could have toxicological effects [52]. The rise in the relative organ weights of the pancreas, brain, and lung at 5000 mg kg<sup>-1</sup> ZnO nanoparticles may be largely attributed to the decrease in body weight [53].

### 3.2 Effects on Liver Tissues

After entering the body through any of the various methods, ZnO nanoparticles may function as a key target organ for the liver, which is the main organ of metabolism. The ability of ZnO nanoparticles to induce apoptosis and genotoxicity in human liver cells (HepG2), as well as the underlying molecular mechanisms of its cellular toxicity. Investigations were done on the part that dissolution plays in the toxicity of ZnO nanoparticles [54]. Given that inhalation is the primary method of exposure to ZnO nanoparticles in the workplace, pulmonary toxicity caused by ZnO nanoparticles has come under increased scrutiny. Acute pulmonary inflammation, chronic inflammation, altered metabolisms, histological abnormalities in the lungs, and airway irritation were among

the toxicity outcomes caused by ZnO nanoparticles that were previously documented in vivo investigations [55].

ZnO nanoparticles, regardless of how they enter the body, preferentially accumulate in the liver. ZnO exposure has been linked to hepatic damage, suggesting that the liver, the body's biggest detoxifying organ, is vulnerable to xenobiotic-mediated damage which is shown in Figure (6). ZnO nanoparticles caused increased levels of the enzymes aspartate transaminase (AST), alanine transaminase (ALT), and lactate dehydrogenase (LDH) in rats and mice after a single oral dosage [56]. Alkaline phosphatase (ALP) levels were shown to be increased in studies using a single intraperitoneal dosage of ZnO nanoparticles. When a single dosage of ZnO nanoparticles was given orally to mice [23]. Histopathological changes in the liver have been reported after a single oral dosage of ZnO nanoparticles in mice, including extensive hepatic edema, vacuolization, cellular necrosis, congestion, and fibrosis. glycogen buildup [34].

**Table 1.** Toxicological effects ZnO nanoparticles on the livers, kidneys, and lungs of rats.

| Organs  | Routes           | Effects  | References          |
|---------|------------------|--|---------------------|
| Livers  | Oral, IM, IV, IP | Abnormal rise of blood liver enzymes • AST, ALT, ALP, and LDH and Histopathological changes  | [19,23,34,48,57,58] |
|         | Oral, IV         | causes significant liver enlargement, vacuolization, cellular necrosis, congestion, and glycogen buildup.  | [34,58]             |
|         | Oral             | Low-dose apoptotic liver alterations and single-dose IP localized inflammation   | [23]                |
|         | IM               | The hepatic sinusoid can be partially dilated  | [42]                |
| Kidneys | IP               | Elevated levels of kidney injury marker BUN and creatine phosphokinase   | [35, 59]            |
|         | Oral             | Histopathological alterations were seen that decreased total kidney glutathione levels.  | [58]                |
|         | IP               | Resulting in necrosis, edema, and hydropic degeneration  | [16]                |
|         | SC, IP           | causes tubular dilation, Focal interstitial edema, and inflammation  | [23]                |
| Lungs   | IP               | Markers for oxidative stress and inflammation were found to be elevated, including lipid peroxide, heme oxygenase-1 (HO-1), and -tocopherol in the lungs       | [60]                |
|         | IT               | Extreme alveolar desquamation caused by massive acute lung inflammation caused   | [61]                |
|         | IT               |  |                     |
|         | IP, IT           | lung and systemic inflammation, dyslipidemia, and elevated blood HO-1 levels.  | [20]                |
|         | IV, IT           | Induces edema, lymphoid cell infiltration, and increased bronchiole epithelial cell proliferation and hypertrophy, induced pulmonary fibrosis and inflammation | [19]                |
|         | IP               | can cause aortic damage; cause serious inflammation, significant hyperemia in the alveoli, and edema; causes mild interstitial inflammation.                   | [20,58]             |
|         | IV               |  | [23]                |
|         | [42]             |  |                     |

System localized infarction at high dosages and early apoptotic changes in the liver were seen after a single intraperitoneal injection of ZnO nanoparticles in rats. We found that 1 day after a single intravenous injection (0.2 mg kg<sup>-1</sup> of ZnO nanoparticles in mice, the hepatic sinusoid was slightly dilated and the Zn concentration in liver was 10.1-8.6 g/g [39].

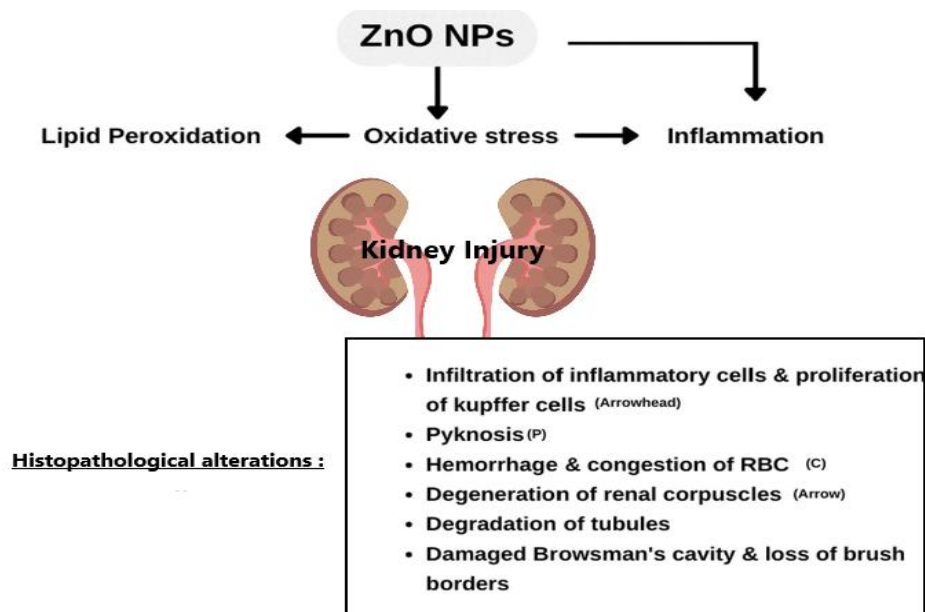
### 3.3 Effects on Kidney Tissues

Yan G et al. [62] examined the biochemical compositions of urine and kidney samples from rats that received doses of 100,

300, and 1000 mg kg<sup>-1</sup> of ZnO nanoparticles over a 14-day period, using a 1H nuclear magnetic resonance (NMR) technique. The results show that ZnO nanoparticles can disrupt energy metabolism and result in mitochondrial and cell membrane impairment in the rat kidney, which may contribute to ZnO nanoparticles-induced nephrotoxicity.

Rani V et al. [63] treated the rats with ZnO nanoparticles (50 mg kg<sup>-1</sup>). Their histopathological studies also showed that the morphology of the liver cells had improved. ZnO nanoparticles may provide protection by selectively intoxicating proliferating





**Figure 7.** Flowchart depicting kidney damage caused by ZnO nanoparticles [51].

tissue, such as the adenomatous islands developed in the liver. The amelioration of DMN-induced toxic effects may also involve zinc metallothionein (Zn-MT), which is induced by ZnO nanoparticles. The major mechanism underlying ZnO nanoparticles' protective properties is still their ability to reduce oxidative stress. ZnO nanoparticles accumulated in the tested organs including kidneys, suggesting that the kidney could be one of the major target organs for the ZnO nanoparticles induced toxicity [35]. The most recent research on the nephrotoxicity caused by ZnO nanoparticles in several animal models. Bowman's gap increases, the distal convoluted tubule is destroyed, there is intratubular protein deposition, inflammatory cells are infiltrated, and there are capillaries clogged between the tubules, among other histological alterations in the kidney. Histological examination of the kidney and serum biochemical analysis [64]. The kidneys, due to their high blood supply and ability to concentrate toxins, are especially susceptible to xenobiotics and are the preferred accumulation site for ZnO nanoparticles following oral ZnO exposure (600 milligrams or 1 gram per kilogram of body weight every day for 5 days) total glutathione in the kidneys is significantly reduced, suggesting functional impairment to kidney tissue [59].

The rats had elevated levels of blood urea nitrogen (BUN) and creatinine (Cre) 6 hours after receiving a single intraperitoneal injection of ZnO nanoparticles (dosage 10 mg kg<sup>-1</sup>). These are biochemical markers of kidney damage and a Zn level of around 70 g/g in the kidneys. In contrast, we found that mice given a single intravenous injection of ZnO nanoparticles (0.2 mg kg<sup>-1</sup>) did not develop any pathological alterations to the kidneys, including an increase in BUN or Cre levels. In comparison, the Zn content in kidneys was 8.6 1.0 g/g [57]. ZnO nanoparticles (size 30 nm, dosage 300 mg kg<sup>-1</sup>) were given orally to mice over the course of 14 days, and Zn content was measured in kidneys to be about 40 g/g, indicating that the tubules had enlarged as a result. After receiving a single intraperitoneal dose of ZnO nanoparticles (size 20 nm, dose 100 g/mL daily for 14 days), the kidneys of mice were inflamed and developed focal interstitial edema, as determined by a pathological investigation. Khorsandi et al. [65] reported that inhaled ZnO nanoparticles cause renal irritation to last for a long time. Malonaldehyde, H<sub>2</sub>O<sub>2</sub>, and NO concentrations in the kidney were reduced in DMN (2 l/100 g body weight/rat)-treated rats when ZnO nanoparticles (50 mg kg<sup>-1</sup> body weight/rat) were administered. The healing of oxidative DNA damage and less apparent histological abnormalities in the

kidney lend weight to these findings. ZnO nanoparticles are thought to be harmful to renal tissue, yet their high therapeutic and antioxidative properties aid in lessening the rat kidney damage caused by dimethylnitrosamine (DMN) [66].

### **3.4 Effects on Lung Tissues**

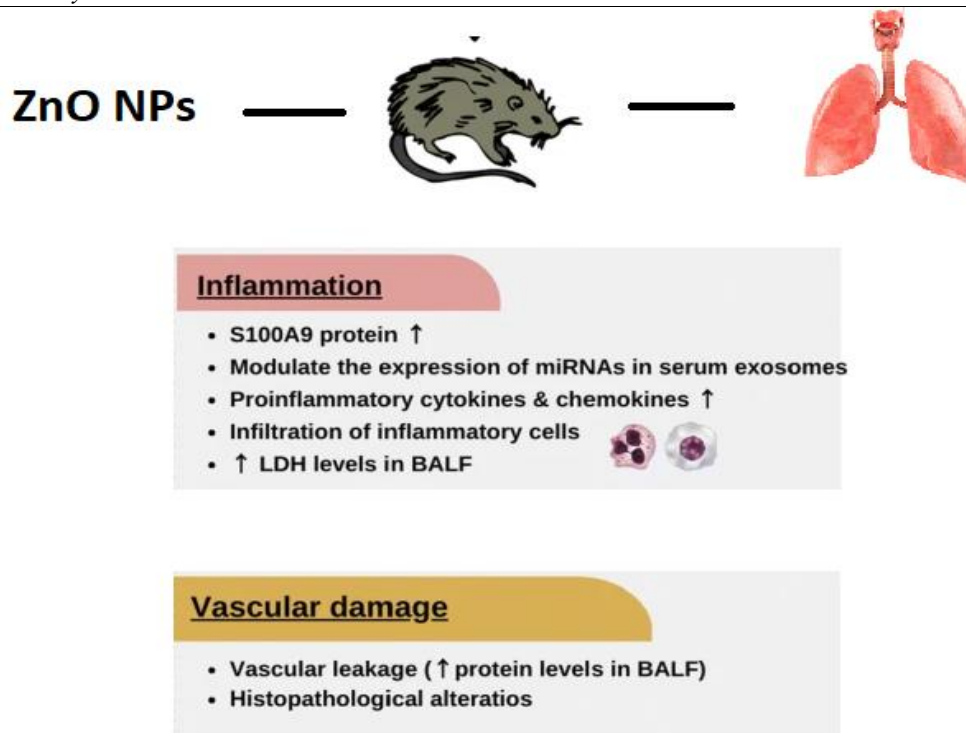
Inhalation is the most common route of ZnO nanoparticles exposure on the job. Studies on the toxicity and injury caused by ZnO nanoparticles to the lungs have mostly involved exposure by inhalation, intratracheal instillation, or intranasal administration. Researchers have shown that inhaling ZnO nanoparticles is the most lethal route of exposure [67]. Due to the acidic nature of lung lining fluid, ZnO nanoparticles degrade and release  $Zn^{+2}$ , leading to ROS-induced inflammation, necrosis, and cell death [68]. High levels of oxidative stress were seen in rats after receiving a single intratracheal injection of ZnO nanoparticles (size 21 nm, dosage  $70 \text{ g mL}^{-1}$ ), as evidenced by an increase in lung lipid peroxide, heme oxygenase-1 ( $HO^{-1}$ ), and -tocopherol. The ZnO nanoparticles also stayed there in the lungs and continued to release  $Zn^{+2}$  [60]. Jacobsen et al. [61] were injected ZnO nanoparticles (size 12–3 nm, low dose– $0.3 \text{ mg kg}^{-1}$ ) into the trachea of mice, and the animals afterward experienced massive acute pulmonary infarction, excessive desquamation of alveolar barrier epithelial cells, and death, and histological alterations (including edema, eosinophilic granuloma, lymphoid cell infiltration, and enhanced proliferation and hypertrophy of bronchiole epithelial cells). An increase in the lung weight to body weight ratio, as well as histological abnormalities such as pulmonary fibrosis and inflammation, were seen 7 days after intratracheal injection of ZnO nanoparticles in mice. Huang et al., studied the effects of ZnO nanoparticles inhalation in mice (dosage  $2.5 \text{ mg/m}^3$ , 5 hours/day for 5 days) and hypothesized that ZnO nanoparticles inhalation may lead to the development of allergic airway inflammation [69]. Figure 5 shows the toxicity effect of in lungs.

Saptarshi et al. [70] investigated the 24 hours inhalation of ZnO nanoparticles (size 30 nm, dose  $5 \text{ mg kg}^{-1}$ ), mice showed

signs of pulmonary inflammation as evidenced by an increase in eotaxin mRNA expression in lung tissue and the release of pro-inflammatory cytokines in their blood. Histopathological abnormalities in the lungs of mice were seen after a single oral exposure to ZnO nanoparticles. These abnormalities included severe inflammation, vascular damage figure (8) severe hyperemia in the alveoli, and edema [71]. Cho W-S et al. [72] administered ZnO nanoparticles intratracheally to rats at two different dosages (50 and  $150 \text{ cm}^2/\text{rat}$ ). They conducted assessments at 24 hours, one week, and four weeks to evaluate the dose-dependent and time-dependent responses. Eosinophilia, airway epithelial cell proliferation, goblet cell hyperplasia, and lung fibrosis were all brought on by ZnO nanoparticles. Chronic bronchocentric interstitial lung fibrosis was linked to elevated myofibroblast accumulation and positive transforming growth factor. The fundamental source of ZnO nanoparticles-induced various progressive severe lung damage is a pH-dependent breakdown of ZnO nanoparticles inside phagosomes. Wang, D et al. [19] were given varied doses of ZnO nanoparticles (200, 400,  $800 \text{ g/kg}$ ) to mice. Animal mortality, organ/body weight ratios, hematological, blood biochemistry, and histopathology were used to determine the acute toxicity of the substance. Malondialdehyde levels in the lung homogenates also rose. In addition, it was found that the lungs had undergone inflammatory and hyperplastic alterations.

### **4. Therapeutic Mechanism of ZnO Nanoparticles**

The exact toxicological mechanisms of ZnO nanoparticles are yet unclear. Nanoparticles have various physical and chemical characteristics, which may be related to their potential toxicity because their surface area is proportionately bigger than that of larger particles [73]. When ingested, ZnO nanoparticles break down and release  $Zn^{+2}$ . Some research has put forward the theory that  $Zn^{+2}$  is responsible for toxicity effects of ZnO nanoparticles [74]. Fukui et al., injected intratracheally ZnO nanoparticles stayed in the lungs of rats, where they constantly produced  $Zn^{+2}$  and caused severe oxidative stress. ZnO nanoparticles induced a rise in 8-hydroxydeoxyguanosine (8-OHdG), a major ROS product and a widely used marker for



**Figure 8.** Lung, vascular damage, and histopathological alterations induced by ZnO nanoparticles [51].

oxidative DNA damage, whether injected intratracheally or breathed. ZnO nanoparticles are capable of producing high amounts of free radicals, which can lead to oxidative damage [60]. Li YS et al. [75] found that 8-OHdG was highly accumulated in the lungs after intratracheal installation of ZnO nanoparticles containing lipopolysaccharides. Possible involvement of oxidative stress in inflammation; this can lead to DNA damage and cell death, or apoptosis. They postulated further that ZnO nanoparticles may cross the blood-air barrier and harm the liver in this way. Previous research examined how much 8-OHdG was excreted in the urine following a single intravenous dose of ZnO nanoparticles. The concentration was significantly higher after day one and decreased dose-dependently over the next six days. Serum superoxide dismutase levels were considerably elevated at 24 and 48 hours after intravenous injection of 0.2 mg kg<sup>-1</sup> ZnO nanoparticles. An in vitro study found that both ZnO nanoparticles and Zn<sup>+2</sup> entered cells. Zn<sup>+2</sup> impacts enzyme balance, transcription factors, and signaling pathways, whereas nanoparticles induce cell inactivation, oxidative stress, mitochondrial damage, and intracellular Ca<sup>+2</sup> excess.

Comparing the effects of ZnO nanoparticles and bulk ZnO on astrocytes revealed that both were toxic, but that astrocytes exposed to ZnO nanoparticles had more ROS production and caspase activity than those exposed to bulk ZnO [76]. Tang et al. [77] found that after a week of oral treatment of ZnO Nanoparticles at 100, 300, and 600 mg kg<sup>-1</sup>, mRNA expression for cytochrome P450 1A2 (CYP1A2) was downregulated, whereas expression for cytochrome P450 2C11 and CYP3A4 was upregulated, and pathological abnormalities were observed in liver and kidney tissues.

## 5. Conclusion

ZnO nanoparticles are rapidly distributed throughout the body and are efficiently eliminated. They are primarily absorbed in ionic form, with some in particle form. Importantly, these nanoparticles do not tend to accumulate in tissues over an extended period. Regardless of the exposure method, higher concentrations of Zn were detected in the key target organs for ZnO nanoparticles, including the liver, kidneys, and lungs. The liver is the primary site of accumulation for ZnO nanoparticles, and exposure through various routes led to histological changes and liver damage. Following a single oral or intraperitoneal

dose, kidney injury was observed. Notably, Lung damage was assessed using intratracheal instillation and inhalation exposure methods. The primary toxicological mechanism associated with ZnO nanoparticles involves the generation of substantial oxidative stress, characterized by the production of significant levels of reactive oxygen species (ROS). ROS generation is attributed to two main factors: the release of Zn<sup>+2</sup> ions from ZnO nanoparticles and the particulate effect resulting from the semiconductor or electronic properties of ZnO nanoparticles. One effective strategy to mitigate the toxicity of these particles is by coating the surface of ZnO nanoparticles with silica, which effectively suppresses ROS production and Zn<sup>+2</sup> ion release.

#### **Data Availability statement**

The data used to support the findings of this study are available from the corresponding author upon request.

#### **Conflicts of Interest**

All authors declare that, they have no conflict of interest.

#### **Author Contributions**

All authors participated in the initial draft creation, reviewed the manuscript, and contributed to the editing process.

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#### **REFERENCES**

1. Haleem, A., Javaid, M., Singh, R. P., Rab, S. and Suman, R., 2023. Applications of nanotechnology in medical field. *Global Health Journal*, 7, pp. 70-77
2. Moshed, A., Sarkar, M. K. I. and Khaleque, M. A., 2017. The application of nanotechnology in medical sciences: New horizon of treatment. *Am. J. Biomed. Sci*, 9, pp. 1-14.
3. Klymchenko, A. S., Liu, F., Collot, M. and Anton, N., 2021. Dye-loaded nanoemulsions: Biomimetic fluorescent nanocarriers for bioimaging and nanomedicine. *Advanced healthcare materials*, 10, pp. 2001289.
4. Kumar, R., Kumar, M. and Luthra, G., 2023. Fundamental approaches and applications of nanotechnology: A mini review. *Materials Today: Proceedings*.
5. Paulkumar, K., Rajeshkumar, S., Gnanajobitha, G., Vanaja, M., Malarkodi, C. and Annadurai, G., 2013. Biosynthesis of silver chloride nanoparticles using *Bacillus subtilis* MTCC 3053 and assessment of its antifungal activity. *International Scholarly Research Notices*, pp. 1-8.
6. Paulkumar, K., Gnanajobitha, G., Vanaja, M., Rajeshkumar, S., Malarkodi, C., Pandian, K. and Annadurai, G., 2014. Piper nigrum leaf and stem assisted green synthesis of silver nanoparticles and evaluation of its antibacterial activity against agricultural plant pathogens. *The Scientific World Journal*, pp. 829-894.
7. Rajeshkumar, S., Malarkodi, C., Paulkumar, K., Vanaja, M., Gnanajobitha, G. and Annadurai, G., 2014. Algae mediated green fabrication of silver nanoparticles and examination of its antifungal activity against clinical pathogens. *International journal of Metals*, pp. 1-8.
8. Subramaniam, V. D., Prasad, S. V., Banerjee, A., Gopinath, M., Murugesan, R., Marotta, F., Sun, X.-F. and Pathak, S., 2019. Health hazards of nanoparticles: Understanding the toxicity mechanism of nanosized ZnO in cosmetic products. *Drug and chemical toxicology*, 42, pp. 84-93.
9. Senthilkumar, K., Senthilkumar, O., Yamauchi, K., Sato, M., Morito, S., Ohba, T., Nakamura, M. and Fujita, Y., 2009. Preparation of ZnO nanoparticles for bio-imaging applications. *Physica status solidi (b)*, 246, pp. 885-888.
10. Madhumitha, G., Elango, G. and Roopan, S. M., 2016. Biotechnological aspects of ZnO nanoparticles: Overview on synthesis and its applications. *Applied microbiology and biotechnology*, 100, pp. 571-581.
11. Poulsen, H. D., 1995. Zinc oxide for weanling piglets. *Acta Agriculturae Scandinavica A-Animal Sciences*, 45, pp. 159-167.
12. Burrough, E. R., De Mille, C. and Gabler, N. K., 2019. Zinc overload in weaned pigs: Tissue accumulation, pathology, and growth impacts. *Journal of Veterinary Diagnostic Investigation*, 31, pp. 537-545.
13. Komatsu, T., Sugie, K., Inukai, N., Eguchi, O., Oyamada, T., Sawada, H., Yamanaka, N. and Shibahara, T., 2020. Chronic pancreatitis in farmed pigs fed excessive zinc oxide. *Journal of Veterinary Diagnostic Investigation*, 32, pp. 689-694.
14. Bonetti, A., Tugnoli, B., Piva, A. and Grilli, E., 2021. Towards zero zinc oxide: Feeding strategies to manage post-weaning diarrhea in piglets. *Animals*, 11, pp. 642.
15. Choi, S.-J. and Choy, J.-H., 2014. Biokinetics of zinc oxide nanoparticles: Toxicokinetics, biological fates, and protein interaction. *International journal of nanomedicine*, 9, pp. 261-269.
16. Sharma, V., Singh, P., Pandey, A. K. and Dhawan, A., 2012. Induction of oxidative stress, DNA damage and apoptosis in mouse liver after sub-acute oral exposure to zinc oxide nanoparticles. *Mutation Research/Genetic Toxicology and Environmental Mutagenesis*, 745, pp. 84-91.

17. Liu, N., Mu, Y., Chen, Y., Sun, H., Han, S., Wang, M., Wang, H., Li, Y., Xu, Q. and Huang, P., 2013. Degradation of aqueous synthesized cdte/zns quantum dots in mice: Differential blood kinetics and biodistribution of cadmium and tellurium. *Particle and fibre toxicology*, 10, pp. 1-9.
18. Baek, M., Chung, H.E., Yu, J., Lee, J.A., Kim, T.H., OH, J.M., Lee, W.J., Paek, S.M., Lee, J. K. and Jeong, J., 2012. Pharmacokinetics, tissue distribution, and excretion of zinc oxide nanoparticles. *International journal of nanomedicine*, pp. 3081-3097.
19. Wang, D., Li, H., Liu, Z., Zhou, J. and Zhang, T., 2017. Acute toxicological effects of zinc oxide nanoparticles in mice after intratracheal instillation. *International journal of occupational and environmental health*, 23, pp. 11-19.
20. Yan, Z., Wang, W., Wu, Y., Wang, W., Li, B., Liang, N. and Wu, W., 2017. Zinc oxide nanoparticle-induced atherosclerotic alterations in vitro and in vivo. *International journal of nanomedicine*, 12, pp. 4433-4442.
21. Holmes, A. M., Kempson, I., Turnbull, T., Paterson, D. and Roberts, M. S., 2020. Penetration of zinc into human skin after topical application of nano zinc oxide used in commercial sunscreen formulations. *ACS Applied Bio Materials*, 3, pp. 3640-3647.
22. Xiong, H. M., 2013. ZnO nanoparticles applied to bioimaging and drug delivery. *Advanced Materials*, 25, pp. 5329-5335.
23. Hong, T.-K., Tripathy, N., Son, H.-J., Ha, K.-T., Jeong, H.-S. and Hahn, Y.-B., 2013. A comprehensive in vitro and in vivo study of ZnO nanoparticles toxicity. *Journal of Materials Chemistry B*, 1, pp. 2985-2992.
24. Jha, A. K., Prasad, K. and Kulkarni, A., 2007. Microbe-mediated nanotransformation: Cadmium. *Nano*, 2, pp. 239-242.
25. Nagajyothi, P., Sreekanth, T., Tettey, C. O., Jun, Y. I. and Mook, S. H., 2014. Characterization, antibacterial, antioxidant, and cytotoxic activities of ZnO nanoparticles using *Coptidis rhizoma*. *Bioorganic & medicinal chemistry letters*, 24, pp. 4298-4303.
26. Hameed, A. S. H., Karthikeyan, C., Ahamed, A. P., Thajuddin, N., Alharbi, N. S., Alharbi, S. A. and Ravi, G., 2016. In vitro antibacterial activity of ZnO and nd doped zno nanoparticles against esbl producing *Escherichia coli* and *Klebsiella pneumoniae*. *Scientific reports*, 6, pp. 24312.
27. Ma, H., Williams, P. L. and Diamond, S. A., 2013. Ecotoxicity of manufactured ZnO nanoparticles—a review. *Environmental pollution*, 172, pp. 76-85.
28. Vimala, K., Sundarraj, S., Paulpandi, M., Vengatesan, S. and Kannan, S., 2014. Green synthesized doxorubicin loaded zinc oxide nanoparticles regulates the bax and Bcl-2 expression in breast and colon carcinoma. *Process biochemistry*, 49, pp. 160-172.
29. Venkatachalam, P., Jayaraj, M., Manikandan, R., Geetha, N., Rene, E. R., Sharma, N. and Sahi, S., 2017. Zinc oxide nanoparticles (ZnONPs) alleviate heavy metal-induced toxicity in *Leucaena leucocephala* seedlings: A physiochemical analysis. *Plant Physiology and Biochemistry*, 110, pp. 59-69.
30. Hazra, C., Kundu, D., Chaudhari, A. and Jana, T., 2013. Biogenic synthesis, characterization, toxicity and photocatalysis of zinc sulfide nanoparticles using rhamnolipids from *Pseudomonas aeruginosa* BS01 as capping and stabilizing agent. *Journal of Chemical Technology & Biotechnology*, 88, pp. 1039-1048.
31. Rajeshkumar, S., 2016. Synthesis of silver nanoparticles using fresh bark of *Pongamia pinnata* and characterization of its antibacterial activity against gram positive and gram negative pathogens. *Resource-Efficient Technologies*, 2, pp. 30-35.
32. Viswanath, B. and Kim, S., 2017. Influence of nanotoxicity on human health and environment: The alternative strategies. *Reviews of Environmental Contamination and Toxicology Volume 242*, pp. 61-104.
33. Amde, M., Liu, J.-f., Tan, Z.-Q. and Bekana, D., 2017. Transformation and bioavailability of metal oxide nanoparticles in aquatic and terrestrial environments. A review. *Environmental pollution*, 230, pp. 250-267.
34. Li, C.H., Shen, C.C., Cheng, Y.W., Huang, S.H., Wu, C.C., Kao, C.C., Liao, J.W. and Kang, J.J., 2012. Organ biodistribution, clearance, and genotoxicity of orally administered zinc oxide nanoparticles in mice. *Nanotoxicology*, 6, pp. 746-756.
35. Lin, Y.-F., Chiu, I.-J., Cheng, F.-Y., Lee, Y.-H., Wang, Y.-J., Hsu, Y.-H. and Chiu, H.-W., 2015. The role of hypoxia-inducible factor-1 $\alpha$  in zinc oxide nanoparticle-induced nephrotoxicity in vitro and in vivo. *Particle and fibre toxicology*, 13, pp. 1-14.
36. Amara, S., Slama, I. B., Mrad, I., Rihane, N., Khemissi, W., El Mir, L., Rhouma, K. B., Abdelmelek, H. and Sakly, M., 2014. Effects of zinc oxide nanoparticles and/or zinc chloride on biochemical parameters and mineral levels in rat liver and kidney. *Human & experimental toxicology*, 33, pp. 1150-1157.
37. Elshama, S. S., El-Kenawy, A. E.-M. and Osman, H.-E. H., 2017. Histopathological study of zinc oxide nanoparticle-induced neurotoxicity in rats. *Toxicology*, 13, pp. 95-103.
38. Choi, J., Kim, H., Kim, P., Jo, E., Kim, H.-M., Lee, M.-Y., Jin, S. M. and Park, K., 2015. Toxicity of zinc oxide nanoparticles in rats treated by two different routes: Single intravenous injection and single oral administration. *Journal of Toxicology and Environmental Health, Part A*, 78, pp. 226-243.
39. Pasupuleti, S., Alapati, S., Ganapathy, S., Anumolu, G., Pully, N. R. and Prakhya, B. M., 2012. Toxicity of zinc oxide nanoparticles through oral route. *Toxicology and Industrial Health*, 28, pp. 675-686.
40. Lee, J., Yu, W.-J., Song, J., Sung, C., Jeong, E. J., Han, J.-S., Kim, P., Jo, E., Eom, I. and Kim, H.-M., 2016. Developmental toxicity of intravenously injected zinc oxide nanoparticles in rats. *Archives of pharmacological research*, 39, pp. 1682-1692.
41. Yeh, T.K., Chen, J.-K., Lin, C.-H., Yang, M.-H., Yang, C. S., Chou, F.I., Peir, J.J., Wang, M.Y., Chang, W.H. and Tsai, M.H., 2012. Kinetics and tissue distribution of

- neutron-activated zinc oxide nanoparticles and zinc nitrate in mice: Effects of size and particulate nature. *Nanotechnology*, 23, pp. 085102.
42. Fujihara, J., Tongu, M., Hashimoto, H., Yamada, T., Kimura-Kataoka, K., Yasuda, T., Fujita, Y. and Takeshita, H., 2015. Distribution and toxicity evaluation of ZnO dispersion nanoparticles in single intravenously exposed mice. *The Journal of Medical Investigation*, 62, pp. 45-50.
43. Vysloužil, J., Kulich, P., Zeman, T., Vaculovič, T., Tvrdoňová, M., Mikuška, P., Večeřa, Z., Stráská, J., Moravec, P. and Balcar, V. J., 2020. Subchronic continuous inhalation exposure to zinc oxide nanoparticles induces pulmonary cell response in mice. *Journal of Trace Elements in Medicine and Biology*, 61, pp. 126511.
44. Konduru, N. V., Murdaugh, K. M., Sotiriou, G. A., Donaghey, T. C., Demokritou, P., Brain, J. D. and Molina, R. M., 2014. Bioavailability, distribution and clearance of tracheally-instilled and gavaged uncoated or silica-coated zinc oxide nanoparticles. *Particle and fibre toxicology*, 11, pp. 1-13.
45. Condello, M., De Berardis, B., Ammendolia, M. G., Barone, F., Condello, G., Degan, P. and Meschini, S., 2016. ZnO nanoparticle tracking from uptake to genotoxic damage in human colon carcinoma cells. *Toxicology in Vitro*, 35, pp. 169-179.
46. Fujihara, J. and Nishimoto, N., 2023. Review of zinc oxide nanoparticles: Toxicokinetics, tissue distribution for various exposure routes, toxicological effects, toxicity mechanism in mammals, and an approach for toxicity reduction. *Biological trace element research*, 1, pp. 1-15.
47. Cho, W.S., Kang, B.C., Lee, J. K., Jeong, J., Che, J.H. and Seok, S. H., 2013. Comparative absorption, distribution, and excretion of titanium dioxide and zinc oxide nanoparticles after repeated oral administration. *Particle and fibre toxicology*, 10, pp. 1-9.
48. K Handral, H. and Kelmani R, C., 2018. A comparative in vivo scrutiny of biosynthesized copper and zinc oxide nanoparticles by intraperitoneal and intravenous administration routes in rats. *Nanoscale research letters*, 13, pp. 1-15.
49. Hong, J.S., Park, M.K., Kim, M.S., Lim, J.H., Park, G.J., Maeng, E.H., Shin, J.H., Kim, M.K., Jeong, J. and Park, J.A., 2014. Prenatal development toxicity study of zinc oxide nanoparticles in rats. *International journal of nanomedicine*, 9, pp. 159-171.
50. Jo, E., Seo, G., Kwon, J.-T., Lee, M., cheun Lee, B., Eom, I., Kim, P. and Choi, K., 2013. Exposure to zinc oxide nanoparticles affects reproductive development and biodistribution in offspring rats. *The Journal of toxicological sciences*, 38, pp. 525-530.
51. Wang, C., Lu, J., Zhou, L., Li, J., Xu, J., Li, W., Zhang, L., Zhong, X. and Wang, T., 2016. Effects of long-term exposure to zinc oxide nanoparticles on development, zinc metabolism and biodistribution of minerals (Zn, Fe, Cu, Mn) in mice. *PloS one*, 11, pp. e0164434.
52. Valdiglesias, V., Costa, C., Kiliç, G., Costa, S., Pásaro, E., Laffon, B. and Teixeira, J. P., 2013. Neuronal cytotoxicity and genotoxicity induced by zinc oxide nanoparticles. *Environment international*, 55, pp. 92-100.
53. Sharma, V., Anderson, D. and Dhawan, A., 2012. Zinc oxide nanoparticles induce oxidative DNA damage and ros-triggered mitochondria mediated apoptosis in human liver cells (hepG2). *Apoptosis*, 17, pp. 852-870.
54. Larsen, S. T., Jackson, P., Poulsen, S. S., Levin, M., Jensen, K. A., Wallin, H., Nielsen, G. D. and Koponen, I. K., 2016. Airway irritation, inflammation, and toxicity in mice following inhalation of metal oxide nanoparticles. *Nanotoxicology*, 10, pp. 1254-1262.
55. Smeets, M. A. and Dalton, P. H., 2005. Evaluating the human response to chemicals: Odor, irritation and non-sensory factors. *Environmental Toxicology and Pharmacology*, 19, pp. 581-588.
56. Chong, C. L., Fang, C. M., Pung, S. Y., Ong, C. E., Pung, Y. F., Kong, C. and Pan, Y., 2021. Current updates on the in vivo assessment of zinc oxide nanoparticles toxicity using animal models. *BioNanoScience*, 11, pp. 590-620.
57. Yan, G., Huang, Y., Bu, Q., Lv, L., Deng, P., Zhou, J., Wang, Y., Yang, Y., Liu, Q. and Cen, X., 2012. Zinc oxide nanoparticles cause nephrotoxicity and kidney metabolism alterations in rats. *Journal of Environmental Science and Health, Part A*, 47, pp. 577-588.
58. Fujihara, J., Tongu, M., Hashimoto, H., Fujita, Y., Nishimoto, N., Yasuda, T. and Takeshita, H., 2015. Pro-inflammatory responses and oxidative stress induced by ZnO nanoparticles in vivo following intravenous injection. *Eur Rev Med Pharmacol Sci*, 19, pp. 4920-4926.
59. Esmaellou, M., Moharamnejad, M., Hsankhani, R., Tehrani, A. A. and Maadi, H., 2013. Toxicity of ZnO nanoparticles in healthy adult mice. *Environmental Toxicology and Pharmacology*, 35, pp. 67-71.
60. Lin, Y.-F., Chiu, I.J., Cheng, F.Y., Lee, Y.H., Wang, Y.J., Hsu, Y.H. and Chiu, H.W., 2015. The role of hypoxia-inducible factor-1 $\alpha$  in zinc oxide nanoparticle-induced nephrotoxicity in vitro and in vivo. *Particle and fibre toxicology*, 13, pp. 1-14.
61. Faddah, L. M., Baky, N. A. A., Al-Rasheed, N. M., Al-Rasheed, N. M., Fatani, A. J. and Atteya, M., 2012. Role of quercetin and arginine in ameliorating nano zinc oxide-induced nephrotoxicity in rats. *BMC complementary and alternative medicine*, 12, pp. 1-14.
62. Fukui, H., Horie, M., Endoh, S., Kato, H., Fujita, K., Nishio, K., Komaba, L. K., Maru, J., Miyauhi, A. and Nakamura, A., 2012. Association of zinc ion release and oxidative stress induced by intratracheal instillation of zno nanoparticles to rat lung. *Chemico-biological interactions*, 198, pp. 29-37.
63. Jacobsen, N. R., Stoeger, T., Van Den Brùle, S., Saber, A. T., Beyerle, A., Vietti, G., Mortensen, A., Szarek, J., Budtz, H. C. and Kermanizadeh, A., 2015. Acute and subacute pulmonary toxicity and mortality in mice after intratracheal instillation of ZnO nanoparticles in three laboratories. *Food and Chemical Toxicology*, 85, pp. 84-95.

64. Rani, V., Verma, Y., Rana, K. and Rana, S. V. S., 2018. Zinc oxide nanoparticles inhibit dimethylnitrosamine induced liver injury in rat. *Chemico-biological interactions*, 295, pp. 84-92.
65. Abdel-Aziz, H. O., Hamdan, H. M. and Ragab, E. E., 2018. The histological effects of zinc oxide nanoparticles on the kidney of adult male rabbits. *Sohag Medical Journal*, 22, pp. 297-301.
66. Khorsandi, L., Heidari-Moghadam, A. and Jozi, Z., 2018. Nephrotoxic effects of low-dose zinc oxide nanoparticles in rats. *Journal of Nephropathology*, 7, pp. 158-165.
67. Rani, V., Verma, Y. and Rana, S., 2022. Zinc oxide nanoparticles ameliorate dimethylnitrosamine-induced renal toxicity in rat. *Applied Biochemistry and Biotechnology*, 194, pp. 1-17.
68. Vanderiel, R. and Jong, W., 2012. A review of mammalian toxicity of ZnO nanoparticles. *Nanotechnol. Sci. Appl*, 5, pp. 61-71.
69. Osmond, M. J. and Mccall, M. J., 2010. Zinc oxide nanoparticles in modern sunscreens: An analysis of potential exposure and hazard. *Nanotoxicology*, 4, pp. 15-41.
70. Huang, K.-L., Chang, H.-L., Tsai, F.-M., Lee, Y.-H., Wang, C.-H. and Cheng, T.-J., 2019. The effect of the inhalation of and topical exposure to zinc oxide nanoparticles on airway inflammation in mice. *Toxicology and Applied Pharmacology*, 384, pp. 114787.
71. Liu, M., Yu, X., Chen, Z., Yang, T., Yang, D., Liu, Q., Du, K., Li, B., Wang, Z. and Li, S., 2017. Aptamer selection and applications for breast cancer diagnostics and therapy. *Journal of nanobiotechnology*, 15, pp. 1-16.
72. Cho, W.S., Duffin, R., Howie, S. E., Scotton, C. J., Wallace, W. A., MacNee, W., Bradley, M., Megson, I. L. and Donaldson, K., 2011. Progressive severe lung injury by zinc oxide nanoparticles; the role of Zn<sup>2+</sup> dissolution inside lysosomes. *Particle and fibre toxicology*, 8, pp. 1-16.
73. Meng, Q., Wang, A., Hua, H., Jiang, Y., Wang, Y., Mu, H., Wu, Z. and Sun, K., 2018. Intranasal delivery of hyperzinc a to the brain using lactoferrin-conjugated n-trimethylated chitosan surface-modified plga nanoparticles for treatment of alzheimer's disease. *International journal of nanomedicine*, 13, pp. 705-718.
74. Camaioni, A., Massimiani, M., Lacconi, V., Magrini, A., Salustri, A., Sotiriou, G. A., Singh, D., Bitounis, D., Bocca, B. and Pino, A., 2021. Silica encapsulation of ZnO nanoparticles reduces their toxicity for cumulus cell-oocyte-complex expansion. *Particle and fibre toxicology*, 18, pp. 1-15.
75. Li, Y.S., Ootsuyama, Y., Kawasaki, Y., Morimoto, Y., Higashi, T. and Kawai, K., 2018. Oxidative DNA damage in the rat lung induced by intratracheal instillation and inhalation of nanoparticles. *Journal of Clinical Biochemistry and Nutrition*, 62, pp. 238-241.
76. Sudhakaran, S., Athira, S. and Mohanan, P., 2019. Zinc oxide nanoparticle induced neurotoxic potential upon interaction with primary astrocytes. *Neurotoxicology*, 73, pp. 213-227.
77. Tang, H.Q., Xu, M., Rong, Q., Jin, R.W., Liu, Q.J. and Li, Y.L., 2016. The effect of ZnO nanoparticles on liver function in rats. *International journal of nanomedicine*, 11, pp. 4275-4285.

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